

1 **Behavioural changes in aposematic *Heliconius melpomene* butterflies in response to**  
2 **their predatory bird calls**

3

4 **Sushant Potdar<sup>1</sup>, Madhuri Dinakar<sup>2</sup>, Erica L. Westerman<sup>1</sup>**

6 <sup>1</sup>Department of Biological Sciences, University of Arkansas, Fayetteville AR

7 <sup>2</sup>KB Group of NY, Inc. dba PRIME AE Group of NY, Albany NY

8

9 Running title: Behavioural change in response to predatory bird calls in *Heliconius*  
10 *melpomene* butterfly

11

12 ORCID

13 Sushant Potdar: 0000-0002-6924-4826

14 Madhuri Dinakar: 0000-0002-9713-6320

15 Erica L. Westerman: 0000-0002-3575-8298

16

17

18

19

20

21

22

23 **Abstract:**

24 Prey-predator interactions have resulted in the evolution of many anti-predatory traits.  
25 One of them is the ability for prey to listen to predators and avoid them. Although prey  
26 anti-predatory behavioural responses to predator auditory cues are well described in a  
27 wide range of taxa, studies on whether butterflies change their behaviours in response to  
28 their predatory calls are lacking. *Heliconius* butterflies are unpalatable and form  
29 Müllerian mimicry rings as morphological defence strategies against their bird predators.  
30 Like many other butterflies in the *Nymphalidae* family, *Heliconius* butterflies possess  
31 auditory organs, which are hypothesized to have evolved to assist with predator detection.  
32 Here we test whether *Heliconius melpomene* change their behaviour in response to their  
33 predatory bird calls by observing the behaviour of male and female *H. m. plessini*  
34 exposed to calls of *Heliconius* avian predators: rufous-tailed jacamar, migratory Eastern  
35 kingbird, and resident tropical kingbird. We also exposed them to the calls of the toco  
36 toucan, a frugivorous bird as a control bird call, and an amplified greenhouse background  
37 noise as a noise control. We found that individuals changed their behaviour in response to  
38 jacamar calls only. Males increased their walking and fluttering behaviour, while females  
39 did not change their behaviour during the playback of the jacamar call. Intersexual  
40 behaviours like courtship, copulation, and abdomen lifting did not change in response to  
41 bird calls. Our findings suggest that despite having primary predatory defences like  
42 toxicity and being in a mimicry ring, *H. m. plessini* butterflies changed their behaviour in  
43 response to predator calls. Furthermore, this response was predator specific, as *H. m.*

44 *plesseni* did not respond to either the Eastern kingbird or the tropic kingbird calls. This  
45 suggests that *Heliconius* butterflies may be able to differentiate predatory calls, and  
46 potentially the birds associated with those calls.

47

48 Key words: auditory cues, Lepidoptera, aposematism, toxic, neotropics, bird  
49 vocalizations

50

51 **Highlights:**

- 52 1. Many prey animals change their behaviour in response to their predator's calls.
- 53 2. Whether butterflies alter behaviour in response to bird predator calls is unknown.
- 54 3. We show that *Heliconius melpomene* change behaviour in response to jacamar calls.
- 55 4. Males increased walking and fluttering, but did not alter courting behaviour.
- 56 5. *H. melpomene* did not respond to predatory Eastern kingbird or tropical kingbird calls.

57

58 **Introduction:**

59 Predation is a ubiquitous interspecific interaction in almost all ecosystems and can  
60 be a strong evolutionary force for the emergence and selection of prey anti-predatory  
61 strategies that increase survival (Lind & Cresswell, 2005). Anti-predatory strategies are  
62 widespread in prey animals and can be morphological or behavioural. Morphological  
63 strategies include aposematism, chemical toxicity, and crypsis (Rojas et al., 2019; Vallin  
64 et al., 2006), while behavioural anti-predatory strategies include active evasion of

65 predatory attacks, and behaviours that decrease detection (Palmer & Packer, 2021). Anti-  
66 predatory strategies can also be a combination of both morphological and behavioural  
67 strategies such as the deimatic displays in mountain katydid *Acripeza reticulata* and  
68 swallowtail butterflies (Olofsson et al., 2012; Umbers & Mappes, 2015).

69 The most common anti-predatory strategies are behavioural responses to detection  
70 and active attacks by predators. These behavioural responses help prey escape predation,  
71 either in the absence of morphological defences, or as a combination with morphological  
72 defences, and can be highly variable across species, within species, and between sexes  
73 (Apfelbach et al., 2005; Lind & Cresswell, 2005). Some species, such as desert isopods  
74 (*Hemilepistus reaumuri*), freeze and retreat inside their burrows upon smelling their  
75 predator's scent (Zaguri & Hawlena, 2020); while others, such as male tree lizards  
76 (*Urosaurus ornatus*) actively escape by fleeing after detecting their predators (Thaker et  
77 al., 2009). Anti-predatory behaviours can also differ within species in response to  
78 different predators, as illustrated by red squirrels (*Tamiasciurus hudsonicus*), which have  
79 different alarm calls for avian predators and ground predators (Greene & Meagher, 1998).  
80 Anti-predatory behavioural responses can also be sex-specific, either due to the inherent  
81 sex-specific differences in physiology and behaviour, or due to the increased vulnerability  
82 of predation during intraspecific sexual behaviours (Curlis et al., 2016; Edomwande &  
83 Barbosa, 2020; Lea & Blumstein, 2011; Sitvarin & Rypstra, 2012; Wormington &  
84 Juliano, 2014). Both males and females are known to alter their courtship and mating  
85 behaviours under predation risk (Acharya & McNeil, 1998; Torsekar et al., 2019). In wolf

86 spider (*Schizocosa ocreata*), males cease courtship behaviour after detecting predatory  
87 birds' calls and take longer to return to courting compared to non-threatening control  
88 sounds (Lohrey et al., 2009), while female túngara frogs (*Physalaemus pustulosus*)  
89 approach calling males more cautiously when exposed to bat wingbeat sounds (Bernal et  
90 al., 2007). The cost of predation during intraspecific sexual behaviours such as courtship  
91 and copulation is high, forcing individuals to switch from sexually oriented behaviours to  
92 survival behaviours.

93 Detecting and recognizing predatory cues are necessary for active predator  
94 avoidance behaviours, and these cues can either be visual, chemical, vibrational, or  
95 auditory. Auditory cues play a vital role in predator avoidance by prey animals, from  
96 invertebrates (Faure & Hoy, 2000; Jacobs et al., 2008; Lohrey et al., 2009; Prakash et al.,  
97 2021; Rosen et al., 2009; Triplehorn et al., 2008) to vertebrates (Bernal et al., 2007;  
98 Cantwell & Forrest, 2013; Deecke et al., 2002). In Lepidoptera (moths and butterflies),  
99 anti-predatory behaviours in moths to predators' auditory cues have been extensively  
100 studied under various ecological contexts. Moths have evolved hearing to detect bat  
101 echolocation calls and avoid bat predation by performing aerial manoeuvres and  
102 jamming echolocation calls (Conner & Corcoran, 2012). Both male and female moths  
103 also reduce sexual activity under bat predation pressure (Acharya & McNeil, 1998;  
104 Edomwande & Barbosa, 2020).

105 While moths are particularly well known for their hearing ability and anti-predator  
106 behaviours, butterflies, their day-flying relatives, are also known to have auditory organs,

107 which may be sensitive to predator sounds (Lane et al., 2008). In particular, many species  
108 in the family *Nymphalidae* possess auditory organs on their wings, such as the blue  
109 morpho *Morpho peleides* (Lane et al., 2008; Lucas et al., 2009; Mikhail et al., 2018),  
110 common wood nymph *Cercyonis pegala* (Sun et al., 2018), the owl butterfly *Caligo*  
111 *eurilochus* (Lucas et al., 2014), butterflies from the genus *Erebia* (Ribarič & Gogala,  
112 1996), and *Heliconius* butterflies (Swihart, 1967). However, unlike moths, it is generally  
113 unknown whether butterflies that possess auditory organs change their behaviour in  
114 response to their predator's vocalizations. In this study, we used a butterfly from the  
115 genus *Heliconius* to test whether these butterflies change their behaviour in response to  
116 their predator's vocalizations.

117 *Heliconius* butterflies (Family Nymphalidae), found in North, Central, and South  
118 America, are toxic, unpalatable, display aposematic colouration, form Müllerian mimicry  
119 rings, and roost communally to avoid bird and bat predation (Engler-Chaouat & Gilbert,  
120 2007; Finkbeiner et al., 2012; Mallet & Gilbert, 1995; Pinheiro De Castro et al., 2019).  
121 Despite these anti-predatory strategies, *Heliconius* butterflies are vulnerable to predation  
122 by specialist bird predators, as well as by naïve generalist predatory birds; and their  
123 mortality is higher when young birds are learning which butterfly species are toxic and  
124 should be avoided (Chai, 1986; Langham, 2004, 2006; Pinheiro, 1996; Pinheiro & Cintra,  
125 2017). Hence, it may be evolutionarily advantageous for *Heliconius* butterflies to detect  
126 the presence of their bird predators and change their behaviours to reduce detection,  
127 despite having multiple anti-predatory strategies. One possible way these butterflies

128 could detect the presence of their bird predators is by using avian vocalization cues,  
129 which are often species specific (Lane et al., 2008; Lucas et al., 2009; Mikhail et al.,  
130 2018). In *Heliconius* butterflies, hearing organs located at the base of the hindwing with  
131 peak sensitivity between 0.5 to 4 KHz at 70-90 dB pressure have been described (Swihart  
132 1967). However, the hypothesis that *Heliconius* butterflies change their behaviour in  
133 response to their predatory birds' vocalizations has never been tested.

134 In this study, we tested whether *Heliconius melpomene plessini* butterflies change  
135 their behaviour in response to the vocalizations of their known bird predators. We first  
136 tested butterfly response to the vocalizations of two predatory birds with disparate calls  
137 as well as the vocalization of a frugivorous bird, to assess whether *H. m. plesseni*  
138 butterflies respond to both predator bird calls and calls of non-predatory birds. After  
139 answering that question, we then tested the response of *H. m. plessini* butterflies to  
140 predators that differ in annual patterns of predation (year-round resident or migratory), to  
141 assess whether strength of *H. m. plesseni* response is associated with degree of annual  
142 avian predator exposure. During both these experiments, we also tested whether  
143 intraspecific sexual behaviours like male courtship and female acceptance/rejection  
144 behaviours changed in response to *H. m. plessini*'s bird predatory calls.

145

## 146 **Materials and Methods**

### 147 *Study species husbandry*

148 *Heliconius melpomene* (Order: *Lepidoptera*, Family: *Nymphalidae*), is native to  
149 Central and South America. The subspecies *H. m. plessini* is found in the mountainous  
150 forests of Ecuador and Peru in South America (Hines et al., 2011). Live pupae of *H. m.*  
151 *plessini* were shipped from Ecodecision Heliconius Works in Quito, Ecuador to the  
152 University of Arkansas Biology greenhouse facility in Fayetteville AR, USA, where they  
153 were maintained at an average temperature of 27°C, average relative humidity of 70%  
154 and a 13:11 hour L:D cycle, to mimic summer tropical conditions. All pupae were hung  
155 and housed in mesh BioQuip cages (34.29 x 34.29 x 60.96 cm, Rancho Dominguez, CA,  
156 U.S.A.) until their eclosion in the greenhouse facility. Newly eclosed individuals were  
157 sexed and tagged with a unique number with a silver metallic permanent marker  
158 (SHARPIE 39108PP) and placed in sex-specific mesh BioQuip cages (60.96 x 60.96 x  
159 142.24 cm) with *ad libitum* BIRDS choice butterfly nectar (Birdschoice, Chilton, WI,  
160 USA) and pollen from *Lantana spp* flowers. Marking butterflies with a marker does not  
161 have long term effects on their behaviour and lifespan (Gall, 1984). Female *H. m. plessini*  
162 were housed with females of two other subspecies, *H. m. malleti* and *H. m. rosina* while  
163 male *H. m. plessini* were housed on their own. Both the male and female cages were  
164 visually isolated from the opposite sex and had no more than 15 individuals in each sex  
165 specific cage at any point in time.

166

167 ***Bird calls and control treatments***



168 We used the calls of four different bird species during our experiments: three  
169 *Heliconius* predators and one frugivore as a control species. Our predatory bird species  
170 were the rufous-tailed jacamar (*Galbula ruficauda*), Eastern kingbird (*Tyrannus*  
171 *tyrannus*), and tropical kingbird (*Tyrannus melancholicus*) (Pinheiro, 1996, 2011;  
172 Pinheiro & Cintra, 2017). We used the non-predatory toco toucan (*Ramphastos toco*) call  
173 to test if *H. m. plessini* respond to bird calls in general, and amplified greenhouse  
174 background noise as a random noise control. We chose toco toucan as our control bird  
175 call because it is a non-predatory frugivorous bird found in the same habitat as our focal  
176 butterflies and has a naturally loud call. Playback recordings of the four bird calls  
177 (rufous-tailed jacamar, Eastern kingbird, tropical kingbird and toco toucan) with minimal  
178 disturbance from background animals were downloaded from Xeno-Canto (Xeno-Canto  
179 Foundation; [www.xeno-canto.org](http://www.xeno-canto.org)) (for sonograms of all calls see Supplementary Figure  
180 2). These bird calls were characterized as ‘songs’ in the original files uploaded on Xeno-  
181 Canto. All the bird calls contain elements within previously reported *Heliconius* hearing  
182 frequency 1-4 KHz (Swihart, 1967), though the main components of the kingbird calls’  
183 are just outside that range at 5 KHz (Supplementary Figure 2).

184 The University of Arkansas butterfly facility has constant and continuous noise  
185 generated by fans and misters which were measured at 65 dB near the behavioural watch  
186 cage using an android sound meter application (Sound Meter-Decibel and noise Meter).  
187 To account for any butterfly behavioural responses to this background noise, or to loud  
188 noises in general, we recorded the greenhouse noise using the android voice recorder

189 application (Voice Recorder, version 3 (42.0)) and used this recording in behavioural  
190 assays as a greenhouse background noise control. During the behavioural assays, the calls  
191 of rufous-tailed jacamar (76 dB), Eastern kingbird (79 dB), tropical kingbird (80 dB),  
192 toco toucan (80 dB) and the greenhouse background noise control (77 dB) were played at  
193 10-15 dB louder than the actual greenhouse background noise. Bird calls in forests are  
194 always against a naturally generated background noise (by other animals; leaves rustling,  
195 waterfalls, and streams). While our constant greenhouse background noise is admittedly  
196 different from that of a forest, the presence of background noise broadly emulates such  
197 sounds generated in the forest. All calls were standardized to one minute long .mp3 files.  
198

### 199 ***Behavioural Assays***

200 All behavioural assays were conducted between 11:00 AM and 2:00 PM, when *H.*  
201 *melpomene* are most active in our greenhouse (Rather et al., 2022). We conducted  
202 behavioural assays using 3-15-day-old males and females in a large behavioural cage  
203 (60.96 x 60.96 x 142.24 cm). In each assay, we used one male and one female and  
204 acclimated them in the behavioural cage for 15 minutes with a JBL® Flip 4 portable blue-  
205 tooth speaker (Harman) and a *Lantana spp.* plant. We used both a male and a female in  
206 our behavioural assay to determine whether predatory bird calls had an effect on  
207 intersexual behaviours (*courtship, abdomen lifting, copulation and, sitting near*) in  
208 addition to any other types of behaviour (*wing fluttering, antennae wiggling, basking,*  
209 *flying, resting, walking*). After a 15-minute acclimation period, we recorded all the

210 behaviours performed by the two individuals in the assay for 15 minutes prior to any  
211 playback calls. We then played one of the bird calls or the control greenhouse background  
212 noise using a JBL® Flip 4 portable blue-tooth speaker from the observer's phone (Google  
213 Pixel), placed inside the behavioural cage for 1 minute and recorded the behaviours of the  
214 two individuals during the playback of the call/background noise. After the playback, we  
215 recorded the behaviours of the two individuals for an additional 14 minutes  
216 (Supplementary Figure 1). We recorded the frequency of *fluttering* and *antenna wiggle*  
217 behaviours and the frequency and duration of *basking*, *flying*, *resting*, *walking*, *courtship*,  
218 *copulation*, *abdomen lifting*, and *sitting near each other* behaviours throughout the entire  
219 30-minute observational period.

220 We defined behaviours for *H. m. plessini* as follows: *fluttering*- opening and closing  
221 of wings either while resting or walking; *antenna wiggle*- movement of antennae at 45°  
222 angle in any direction (Robertson et al., 2020); *basking*- individuals sitting with wings  
223 partially or fully open; *flying*- movement from one point to another in the air using rapid  
224 wing flaps; *resting*- individuals sitting with wings fully closed (Rather et al. 2022);  
225 *walking*- movement from one point to another along the substrate using the legs;  
226 *courtship*- sequences of behaviours where males hover, land and rapidly flap their wings  
227 next to females, and bend their abdomen to initiate copulation (Klein & De Araújo,  
228 2010); *copulation*- where both male and female are mating; *abdomen lifting*- raising the  
229 abdomen at an angle from the normal resting body axis, usually performed by females as  
230 a courtship rejection behaviour (Chouteau et al., 2017); *sitting near each other*- where

231 both individuals are resting or basking within one wingspan from each other (Robertson  
232 et al. 2020).

233 We used Spectator Go (BIOOBSERVE, Fort Lee, NJ, USA) software on an Apple  
234 iPad (1<sup>st</sup> generation) to manually record the frequency and duration of behaviours  
235 performed by the two individuals during the assay. This software enables the observer to  
236 record user defined behaviours in real time, separately for the two individuals, without  
237 instantly visualizing quantities during the recording, and has been used in previous  
238 studies to observe and record butterfly behaviours (Rather et al., 2022; Robertson et al.,  
239 2020; Westerman et al., 2014). To reduce observer bias, only one observer recorded all  
240 the behaviours in this study. We did not use a video camera to record behaviours as some  
241 butterfly inter-individual interactions are minute and nuanced happening at a close range,  
242 while others occupy the full three-dimensional flight area of the cage, and simultaneously  
243 capturing both of these types of behaviours is challenging for a stationary camera, but  
244 relatively straightforward for a trained human observer. Within each experiment, we  
245 tested each male-female pair with all calls with at least 24 hours between each call assay,  
246 and randomized the order of calls for each pair. If either of the butterflies in the pair died  
247 between the assays, then those pairs were eliminated from being tested for the remaining  
248 calls.

249

250 ***Experiment 1: Do *H. m. plesseni* butterflies behaviourally respond to predator bird***  
251 ***calls***

252 To test whether *H. m. plessini* butterflies respond to their avian predator calls or to  
253 other birds or loud random noises in general, we subjected the butterflies to four call  
254 treatments in this experiment: rufous-tailed jacamar (N=22 pairs), Eastern kingbirds  
255 (N=22 pairs),\_toco toucan (N=22 pairs) and greenhouse background noise control (N=18  
256 pairs) using the behavioural assay described above with the calls randomized. We  
257 conducted Experiment 1 from February 2019 to March 2020.

258

259 ***Experiment 2: Does predator residence status influence butterfly response to bird call***

260 Due to the results of Experiment 1 (see below), we conducted a follow up  
261 experiment to test whether predator residence status (migratory or present year-round)  
262 influenced likelihood of *H. m. plessini* butterflies changing their behaviour in response to  
263 predator call. For this experiment, we used the calls of the resident tropical kingbird and  
264 the migratory Eastern kingbird, as they have vocalizations in the same auditory  
265 frequencies, and are more closely related than the Eastern kingbird and rufous-tailed  
266 jacamar. We subjected butterflies to three call treatments: resident tropical kingbird  
267 (N=23 pairs), migratory Eastern kingbird (N=22 pairs) and the control greenhouse  
268 background noise control (N=25 pairs) using the same behavioural assay as Experiment  
269 1, as described above. We conducted Experiment 2 from August to December 2021. We  
270 conducted the same statistical analyses for both Experiment 1 and Experiment 2, albeit  
271 separately.

272

273 ***Statistical analyses***

274 We downloaded the data from Spectator Go software and converted them into .csv  
275 files. Each file consisted of approximately 15 minutes of data, and each assay had four  
276 files (15 minutes before, and during plus after call for male and female separately). Each  
277 bout of behaviour was recorded separately by the software for the 10 behaviours  
278 described above. A *de novo* python code (supplementary material 2) was written to add  
279 each bout of a behaviour and provide the total time spent performing that particular  
280 behaviour. This way, we got the total time spent by an individual butterfly performing  
281 behaviours for the whole assay. Further, we manually extracted the behavioural states  
282 before and after the start and end of calls, as well as extracted the behaviours performed a  
283 minute before, during and after the calls. We performed three separate analyses for each  
284 experiment: behavioural state change between before and after the start and end of calls;  
285 short term (1 minute) changes in behaviours between before, during, and after calls; and  
286 long term (14 minutes) changes in behaviour before and after calls.

287 To determine whether butterflies changed their behavioural state in response to bird  
288 call, we compared the behaviours performed across three time points of an assay: 1)  
289 before vs after the start of call; 2) before vs after the end of call; and 3) before start vs  
290 after end of the call. We used generalized linear mixed models (GLMM) with change in  
291 behaviour between the above time points (yes or no) as the response variable, treatment  
292 (calls), and sex (male or female) as fixed predictor variables, and the order of calls as a  
293 random predictor variable. We later used a pairwise Fisher's test to determine if the

294 proportion of individuals that changed their behaviours were similar or different between  
295 the treatments (bird calls and noise control).

296 To test if the frequency and duration of short-term behaviours changed during and  
297 after a call compared to before a call, we extracted the frequency of *fluttering* and  
298 *antenna wiggle* and duration of the other eight behaviours for the minute before, minute  
299 during, and minute after the call. We performed Principal Component Analysis (PCA) for  
300 the behavioural data during these three minutes, to identify the correlation between  
301 different behaviours and identify new composite behavioural variables. We removed  
302 *abdomen lifting* from the male data set and *courtship* from the female data set as males  
303 and females respectively did not perform these behaviours. We fit a linear mixed model  
304 (LMMs), followed by an ANOVA, with treatment (bird call), state (before, during, and  
305 after call) and their interaction as fixed predictor variables, the order of the calls as a  
306 random predictor variable, and the first three principal components as the response  
307 variables. Further, we performed a Tukey HSD test to determine the pairwise differences  
308 between different combinations of treatment (bird call) and state (before, during, and  
309 after call). Later, we tested whether male *courtship*, *sitting near each other*, female  
310 *abdomen lifting*, *copulation* behaviours changed in response to bird calls by fitting  
311 LMMs followed by an ANOVA, with the same predictor variables. We ran these models  
312 for males and females separately, as males and females performed different behaviours.  
313 We also performed these analyses separately for experiments 1 and 2.

314           Next, to test if there was a prolonged long term response of butterfly behaviour to  
315 the bird calls, we extracted the frequency of *fluttering* and *antenna wiggle* and duration of  
316 the other eight behaviours for the 14 minutes before the call and the 14 minutes after the  
317 call, and performed a PCA for these 28 minutes, again removing *abdomen lifting*  
318 behaviour from male data set and *courtship* behaviour from the female data set. We fit  
319 LMM, followed by an ANOVA, with treatment (bird call), state (before, and after call),  
320 and their interaction as the fixed predictor variables, the order of the calls as a random  
321 predictor variable, and the first three principal components as the response variables for  
322 each sex. Further, we performed a Tukey HSD test to determine the pairwise differences  
323 between different combinations of treatment (bird call) and state (before, and after call).  
324 We also tested whether male *courtship*, *sitting near*; female *abdomen lifting*, *copulation*  
325 behaviours changed in response to bird calls by fitting LMMs followed by an ANOVA,  
326 with the same predictor variables. We again ran these models for both males and females  
327 separately, and performed these analyses separately for experiments 1 and 2.

328           All statistical analyses were run using R version 4.3.0 (R Core Team, 2023). All  
329 plots were generated using *ggplot2* (Wickham, 2016) package.

330

### 331 **Ethical Note**

332           All butterflies used in this study were maintained in climate-controlled greenhouse  
333 conditions similar to those of their natural habitat, as stated in the U.S. Department of  
334 Agriculture, Animal and Plant Health Inspection Service permits P526P-17-00343 and



335 P526P-20-00417. Before and after the assays, all butterflies were maintained in cages  
336 with *ad libitum* food (nectar and flowering *Lantana spp.* plants for pollen). After the  
337 assays, they were moved to breeding cages with *ad libitum* food, where they were kept  
338 until natural death. No butterflies were sacrificed for the purpose of this study.

339

#### 340 **Experiment 1 Results:**

341 ***H. m. plessini* immediately changed their behavioural state in response to the rufous-**  
342 **tailed jacamar call**

343 *H. m. plessini* butterflies immediately changed their behavioural state when the  
344 rufous-tailed jacamar call started ( $\chi^2= 16.03$ ,  $p<0.01$ ; Supplementary Figure 3A;  
345 Supplementary Table 1, 2), when the jacamar call stopped ( $\chi^2= 17.47$ ,  $p<0.001$ ;  
346 Supplementary Figure 4A; Supplementary Table 3, 4), and when compared between  
347 before the call started versus after the call ended ( $\chi^2= 27.12$ ,  $p<0.001$ , Table 1, 2, Figure  
348 1A). They did not significantly change their behavioural state in response to any other  
349 bird call, or in response to the noise control (Supplementary Table 1, 2, 3, 4; Table 1, 2).  
350 We did not find an effect of sex on the change in behavioural state when the calls started,  
351 when the calls stopped, or when compared between before the calls started versus after  
352 the calls ended nor was there an effect of call order on butterfly response (Supplementary  
353 Table 1, 3; Table 1).

354

355 ***H. m. plessini* males increased their walking and fluttering behaviour during the**  
356 **playback of the rufous-tailed jacamar call:**

357       When combining the behavioural data for the 3 minutes before, during, and after  
358 each call in a PCA for each sex, male PC2 values were higher during the rufous-tailed  
359 jacamar call compared to before and after the rufous-tailed jacamar call, and compared to  
360 before, during, and after the Eastern kingbird, toco toucan, and greenhouse background  
361 noise (ANOVA,  $F= 2.336$ ,  $Df= 6$ ,  $p= 0.0328$ ; Figure 2C; Table 3; Supplementary Table 7;  
362 see Supplementary Table 5 for PCA loadings). There was no effect of any of the bird calls  
363 or greenhouse background noise control on male PC1 (Figure 2A; Table 3;  
364 Supplementary Table 7, 8), male PC3 (Table 3; Supplementary Table 7), female PC1  
365 (Figure 2B; Table 3; Supplementary Table 7; see Supplementary Table 6 for PCA  
366 loadings), female PC2 (Figure 2D; Table 2; Supplementary Table 7), or female PC3  
367 (Table 2; Supplementary Table 7).

368

369 ***H. m. plessini* males and females had no long-term changes in behaviour in response**  
370 **to calls:**

371       When combining the behavioural data for the 14 minutes before and after each call  
372 in a PCA for each sex, there was no effect of any of the bird calls or greenhouse  
373 background noise control on male PC1 (Supplementary Figure 5A; Supplementary Table  
374 10, 11; see Supplementary Table 8 for PCA loadings), male PC2 (Supplementary Figure  
375 5C; Supplementary Table 10, 11), male PC3 (Supplementary Table 10, 11); female PC1

376 (Supplementary Figure 5B; Supplementary Table 10, 11; see Supplementary Table 9 for  
377 PCA loadings), female PC2 (Supplementary Figure 5D; Supplementary Table 10, 11), or  
378 female PC3 (Supplementary Table 10, 11).

379

### 380 **No effect of predatory bird calls on *H. m. plessini* intersexual behaviours**

381 Male *courtship, sitting near each other, female abdomen lifting, copulation*  
382 behaviours did not have short-term or long-term changes in response to any bird calls  
383 (Table 3; Supplementary Table 10).

384

### 385 **Experiment 2 results:**

386 While there are a number of hypotheses as to why *H. m. plessini* did not change  
387 their behaviour in response to the migratory Eastern kingbird calls, but did change their  
388 behaviours in response to the resident jacamar calls, two we found particularly interesting  
389 were 1) that jacamars are year round residents while Eastern kingbirds are migratory; and  
390 2) jacamars and Eastern kingbirds have different call frequencies (Hz ranges). To test the  
391 hypothesis that residence status is driving *H. m. plesseni* behavioural response while  
392 holding call frequency (Hz) constant, we then tested whether *H. m. plessini* butterflies  
393 changed their behaviour in response to the resident tropical kingbird call compared to the  
394 migratory Eastern kingbird call in Experiment 2, as these two kingbird species have  
395 vocalizations in the same auditory frequency ranges (Supplementary Figure 2).

396

397 **Residence status of kingbirds did not change *H. m. plessini* behavioural state:**

398 We found that *H. m. plessini* butterflies did not change their behavioural state when  
399 either of the resident or migratory kingbird calls or greenhouse background noise started  
400 (Supplementary Figure 3B; Supplementary Table 12, 13), when either of the kingbird  
401 calls or greenhouse background noise stopped (Supplementary Figure 4B, Supplementary  
402 Table 14, 15), and when compared between before the kingbird calls started versus after  
403 the kingbird calls ended, as well as between before the start and after the end of  
404 greenhouse background noise (Figure 1B; Supplementary Table 16, 17). We did not find  
405 an effect of sex on the change in behavioural state when calls started, when the calls  
406 stopped, and when compared between before the calls started versus after the calls ended  
407 nor was there an effect of call order on butterfly response (Supplementary Table 12, 14,  
408 16).

409

410 **Residence status of kingbirds did not change short-term *H. m. plessini* behaviours:**

411 When combining the behavioural data for the 3 minutes before, during, and after  
412 each call in a PCA for each sex, there was no effect of any kingbird calls or greenhouse  
413 background noise control on male PC1 values (Figure 3A; Supplementary Table 20, 21;  
414 see Supplementary Table 18 for PCA loadings), on male PC2 (Figure 3C; Supplementary  
415 Table 20, 21), male PC3 (Supplementary Table 20, 21), female PC1 (Figure 3B;  
416 Supplementary Table 20, 21; see Supplementary Table 19 for PCA loadings), female PC2

417 (Figure 3D; Supplementary Table 20, 21), and female PC3 (Supplementary Table 20, 21)  
418 values.

419

420 **Residence status of kingbirds did not change long-term *H. m. plessini* behaviours:**

421 When combining the behavioural data for the 14 minutes before and after each call  
422 in a PCA for each sex, there was no effect of any of the kingbird calls or greenhouse  
423 background noise control on male PC1 (Supplementary Figure 6A; Supplementary Table  
424 24, 25; see Supplementary Table 22 for PCA loadings), male PC2 (Supplementary Figure  
425 6C; Supplementary Table 24, 25), male PC3 (Supplementary Table 24, 25); female PC1  
426 (Supplementary Figure 6B; Supplementary Table 24, 25; see Supplementary Table 23 for  
427 PCA loadings), female PC2 (Supplementary Figure 6D, Supplementary Table 24, 25), or  
428 female PC3 (Supplementary Table 24, 25).

429

430 **No effect of predatory kingbird calls on *H. m. plessini* intersexual behaviours**

431 Male *courtship, sitting near each other*, female *abdomen lifting, copulation*  
432 behaviours did not have short-term or long-term changes in response to any bird calls  
433 (Supplementary Table 20, 24).

434

435 **Discussion:**

436 *Heliconius melpomene plessini* butterflies changed their behaviour in response to  
437 predatory rufous-tailed jacamar calls but did not change their behaviour in response to

438 predatory Eastern kingbird or tropical kingbird calls. We found a sex-specific difference  
439 in behaviour, where males, but not females, increased their fluttering and walking  
440 behaviours during the playback of the rufous-tailed jacamar calls. The observed  
441 behavioural changes in response to rufous-tailed jacamar calls are short-term and do not  
442 persist over an extended duration of time.

443 A major finding of this study is that toxic, unpalatable, and aposematic *Heliconius*  
444 *melpomene plessini* butterfly changed their behaviour in response to the predatory rufous-  
445 tailed jacamar calls. Contrary to our expectations, *H. m. plessini* butterflies did not  
446 change their behaviour in response to either the Eastern kingbird or tropical kingbird  
447 calls. Two non-mutually exclusive hypotheses can be postulated to explain these results:  
448 1) There may be reduced predation pressure from both the Eastern and tropical kingbirds  
449 compared to rufous-tailed jacamar, which has led to an evolved behavioural response to  
450 the rufous-tailed jacamar but not to two kingbird species, and/or 2) *H. m. plessini* may be  
451 incapable of hearing the Eastern and tropical kingbird calls. Since Eastern kingbirds are  
452 migratory and tropical kingbirds are year-round residents, we had hypothesized that, if  
453 *H.m. plessini* could hear the Eastern and tropical kingbird calls, they might respond to the  
454 resident tropical kingbird due to their year-round presence, but not the migratory Eastern  
455 kingbird. Tropical kingbird calls are similar in frequency to the calls of Eastern kingbird  
456 (Supplementary Figure 2). However, we found that *H. m. plessini* butterflies did not  
457 change their behaviour in response to either of the kingbird calls, suggesting that between

458 kingbird species variation in predation pressure was not sufficient to induce *H. m.*  
459 *plesseni* variation in response to Eastern and tropical kingbird calls.

460 *H. m. plessini* may be under reduced predation pressure from kingbirds relative to  
461 rufous-tailed jacamars. While the rufous-tailed jacamar is a year-round resident of *H. m.*  
462 *plesseni*'s habitat, the Eastern kingbird is migratory and is not present during half of the  
463 year in South America, where *H. m. plessini* is found. Eastern kingbird is also frugivorous  
464 during their migration over Central and South America (Blake & Loiselle, 1992; Morton,  
465 1971). While this does not explain the lack of response to the tropical kingbird, an  
466 additional possibility is that *Heliconius melpomene* may be differentially palatable for  
467 rufous tailed jacamars and tropical kingbirds. Future studies should explore whether there  
468 is variability in toxicity across different subspecies of *H. melpomene*, or variability in  
469 predator sensitivity to *Heliconius* toxicity. Although there is no current support for this  
470 hypothesis in *Heliconius*, the aposematic striped skunks (*Mephitis mephitis*) perform anti-  
471 predatory behaviour in response to the calls of the great horned owl (*Bubo virginianus*)  
472 from which they are not chemically defended, but not in response to the calls of the  
473 coyote (*Canis latrans*), from which they are chemically defended (Fisher & Stankowich,  
474 2018). Moreover, we found that *H. m. plessini* did not change their behaviour in response  
475 to the frugivorous control toucan bird call, despite the toucan calls being in the range  
476 of *Heliconius* hearing, which may indicate that *Heliconius* butterflies are capable of  
477 differentiating between predatory and non-predatory bird calls.

478 An alternative hypothesis is that *H. m. plessini* butterflies may not be capable of  
479 detecting kingbird calls but are able to detect the rufous tailed jacamar calls. Rufous  
480 tailed jacamar calls have a peak frequency below 4 kHz (Mikhail et al. 2018;  
481 Supplementary Figure 2), whereas both the Eastern and tropical kingbirds have a peak  
482 call frequency above 4 kHz (Supplementary Figure 2). Previous electrophysiological tests  
483 of the auditory organ in *H. erato* found that *H. erato* butterflies have the best hearing  
484 capabilities below 4 kHz at 70-90 dB power (Swihart, 1967). Any calls with frequencies  
485 above 4 kHz will require a higher decibel power to hear, which may be the case with the  
486 kingbird calls, as their peak call frequency is between 5-8 kHz. Similar trends have been  
487 observed in the blue morpho (*Morpho peleides*), and common wood nymph (*Cercyonis*  
488 *pegala*) butterflies, where a higher decibel power is required for higher frequency calls to  
489 elicit a response, and that these butterflies are tuned to hear sounds below 5 kHz  
490 (Fournier et al., 2013; Mikhail et al., 2018; Sun et al., 2018). Future studies in *Heliconius*  
491 can test this hypothesis by recording the butterfly responses to reduced frequency (below  
492 4 kHz) kingbird calls and enhanced frequency (above 5kHz) rufous-tailed jacamar calls,  
493 and observe whether *H. m. plesseni* butterflies behaviourally respond to the altered  
494 kingbird and jacamar calls. We also found that *H. m. plessini* did not change their  
495 behaviour in response to the toco toucan calls and greenhouse background noise despite  
496 their calls being below 4 KHz, suggesting that *H. m. plessini* are able to distinguish  
497 between bird calls within their hearing range.



498 Our study is testing the hypothesis that *Heliconius* change their behaviours in  
499 response to predatory bird calls. Although an auditory organ has not yet been described in  
500 *Heliconius melpomene*, the auditory organ is described in a closely related butterfly  
501 *Heliconius erato* (Swihart, 1967). Here we do provide evidence that *H. m. plessini*  
502 changed their behaviour after hearing their predatory rufous-tailed jacamar calls. Future  
503 work can explore the presence of a morphological hearing structure in *Heliconius*  
504 *melpomene plessini* and their electrophysiological range like that performed in other  
505 butterflies (Lane et al. 2008; Lucas et al. 2009; Mikhail et al. 2018), to enhance our  
506 understanding of the physiological mechanisms *H. m. plesseni* may be using to facilitate  
507 their response to the rufous-tailed jacamar.

508 We found that males, but not females, changed their behaviour in response to the  
509 rufous-tailed jacamar calls. This male-specific response to predators is similar to that  
510 found in other species, and may reflect sexual dimorphic predation pressures. Previous  
511 studies in wolf spiders (*Pardosa milvina*) have found that males, but not females, used a  
512 predatory chemical cue experience to decrease predation from a live predator (Sitvarin  
513 and Rypstra, 2012). Similarly, male yellow-billed marmots (*Marmota flaviventris*)  
514 decreased foraging followed by a playback of alarm calls (Lea and Blumstein, 2011). The  
515 sex-specific differences observed in the response of *H. m. plessini* might reflect  
516 differences in predation pressures between the sexes. Male *Heliconius* butterflies in the  
517 wild spend greater time flying in the middle of the forest canopy, and mostly near their  
518 larval/food plants in search of females or for foraging whereas female *Heliconius* spend

519 time fluttering near the understory in search of host plants for egg-laying (Mallet and  
520 Gilbert, 1995). Jacamars and kingbirds are “aerial hawking” predators that catch insects  
521 in flight (Fitzpatrick, 1980), and flying male butterflies might be at greater risk of  
522 predation. This may be the reason for increased fluttering and walking during the jacamar  
523 calls. Moreover, Swihart observed fast wing flutters in *H. erato* when he exposed them to  
524 loudspeaker generated sound (Swihart, 1967), indicating that butterflies may have an  
525 innate wing fluttering response to sound cues. Similar results have been found in *Erebia*  
526 butterflies, where they flutter in response to sound (Ribaric and Gogala 1996) and in the  
527 peacock butterflies (*Inachis io*) where they walk and flutter to avoid rodent predation  
528 during winter hibernation (Olofsson et al., 2011). In *Heliconius*, fluttering may advertise  
529 aposematic colouration and could reinforce the birds’ learned behaviour to avoid brightly  
530 coloured butterflies (Langham, 2006). Similar to the mimicry of aposematic colours  
531 among *Heliconius* species, there is also evidence of locomotor mimicry in the flight of  
532 unpalatable *Heliconius*, including flight measures associated with response to jacamars  
533 (Chai & Srygley, 1990; Srygley, 1994). Future studies of the responses in *H. melpomene*,  
534 their model *H. erato* and other species of the same aposematic mimicry rings could  
535 inform us if certain predators have influenced the evolution of mimetic behavioural  
536 responses.

537 Palatability experiments with jacamars have found that experienced birds sight-  
538 reject flying *Heliconius* butterflies (Pinheiro & Campos, 2019). Therefore, flying,  
539 fluttering and walking behaviours could be advantageous under different ecological

540 contexts (for example bird predator community and experience) as an immediate  
541 response to predator's presence, which may be another reason why we did not see the  
542 behavioural changes over a long-term (14 minutes) period. Future studies could look at  
543 the advantages of these behaviours under different ecological contexts such as  
544 microhabitats (Dell'Aglio et al., 2022), as well as test the behavioural responses of the  
545 butterflies using other predatory birds.

546

#### 547 **Conclusions:**

548 We found that unpalatable and brightly coloured *Heliconius melpomene plessini*  
549 butterflies respond and change their behaviour during the playback of the rufous-tailed  
550 jacamar call. This change in behaviour is sex-specific, where males, but not females,  
551 increase their walking and fluttering behaviour over a short time-frame. Males reverted  
552 back to their original behaviour after the call ended. *H. m. plessini* did not change their  
553 behaviour in response to the two kingbird and the toco toucan calls. Our study opens  
554 avenues for future research in the field of butterfly auditory anti-predatory behaviour  
555 response, its mechanistic underpinnings and ecological and evolutionary consequences,  
556 especially in the context of mimicry.

557

#### 558 **Acknowledgments:**

559 We thank David A. Ernst, Deonna N. Robertson, Grace Hirzel, Matthew Murphy, Yi Ting  
560 Ter, Kiana Kasmaï, and Keity Farfán Pira for their contribution towards *Heliconius*

561 butterfly husbandry and reviewing the manuscript. We thank Brian Counterman for  
562 providing valuable inputs and reviewing this manuscript. We also thank Pooja Panwar for  
563 fruitful discussions during the conceptualization of this project.

564

565 **Data Availability:**

566 Analyses reported in this article can be reproduced using the data provided by the authors  
567 in Dryad (link: XXXX)

568

569 **Supplementary Materials:**

570 Supplementary material 1: a PDF file containing all supplementary figures and tables  
571 (supplementary\_material\_1.pdf)

572 Supplementary material 2: *De novo* python code to extract behavioural data from  
573 SPECTATOR GO! Software files (supplementary\_material\_2.txt)

574 Supplementary material 3: An excel file containing the results of linear mixed models for  
575 (supplementary\_material\_3.xlsx)

576

577 **Funding:**

578 This work was supported by an Arkansas Biosciences Institute grant to ELW, a  
579 Lepidopterists' Society Ron Leuschner Memorial Fund grant to SP, and the University of  
580 Arkansas.

581

582 **CRedit author contributions:**

583 SP- Conceptualization, Methodology, Investigation, Data curation, Formal analysis,  
584 Funding acquisition, Writing- original draft, Writing- review and editing; MD- Data  
585 curation, Software, Writing- review and editing; ELW- Conceptualization, Methodology,  
586 Supervision, Resources, Funding acquisition, Writing- original draft, Writing- review and  
587 editing.

588

589 **References:**

Acharya, L., & McNeil, J. N. (1998). Predation risk and mating behavior: The responses of moths to bat-like ultrasound. *Behavioral Ecology*, *9*(6), 552–558.

<https://doi.org/10.1093/beheco/9.6.552>

Apfelbach, R., Blanchard, C. D., Blanchard, R. J., Hayes, R. A., & McGregor, I. S. (2005). The effects of predator odors in mammalian prey species: A review of field and laboratory studies. *Neuroscience & Biobehavioral Reviews*, *29*(8), 1123–1144. <https://doi.org/10.1016/j.neubiorev.2005.05.005>

Bernal, X. E., Stanley Rand, A., & Ryan, M. J. (2007). Sexual differences in the behavioral response of Túngara Frogs, *Physalaemus pustulosus*, to cues associated with increased predation risk: Sex differences in behavioral response to increased predation risk. *Ethology*, *113*(8), 755–763.

<https://doi.org/10.1111/j.1439-0310.2007.01374.x>

Blake, J. G., & Loiselle, B. A. (1992). Fruits in the diets of neotropical migrant birds in Costa Rica. *Biotropica*, *24*(2), 200. <https://doi.org/10.2307/2388674>

- Cantwell, L. R., & Forrest, T. G. (2013). Response of *Anolis sagrei* to acoustic calls from predatory and nonpredatory birds. *Journal of Herpetology*, 47(2), 293–298.  
<https://doi.org/10.1670/11-184>
- Chai, P. (1986). Field observations and feeding experiments on the responses of rufous-tailed jacamars (*Galbula ruficauda*) to free-flying butterflies in a tropical rainforest. *Biological Journal of the Linnean Society*, 29(3), 161–189.  
<https://doi.org/10.1111/j.1095-8312.1986.tb01772.x>
- Chai, P., & Srygley, R. B. (1990). Predation and the flight, morphology, and temperature of neotropical rain-forest butterflies. *The American Naturalist*, 135(6), 748–765.  
<https://doi.org/10.1086/285072>
- Chouteau, M., Llaurens, V., Piron-Prunier, F., & Joron, M. (2017). Polymorphism at a mimicry supergene maintained by opposing frequency-dependent selection pressures. *Proceedings of the National Academy of Sciences*, 114(31), 8325–8329.  
<https://doi.org/10.1073/pnas.1702482114>
- Conner, W. E., & Corcoran, A. J. (2012). Sound strategies: The 65-Million-Year-Old battle between bats and insects. *Annual Review of Entomology*, 57(1), 21–39.  
<https://doi.org/10.1146/annurev-ento-121510-133537>
- Core Team, R. (2023). *R: A Language and Environment for Statistical Computing*. R Foundation for Statistical Computing. <https://www.R-project.org/>
- Curlis, J. D., Macklem, D. C., Davis, R., & Cox, C. L. (2016). Sex-specific antipredator response to auditory cues in the black spiny-tailed iguana. *Journal of Zoology*, 299(1), 68–74. <https://doi.org/10.1111/jzo.12326>
- Deecke, V. B., Slater, P. J. B., & Ford, J. K. B. (2002). Selective habituation shapes acoustic predator recognition in harbour seals. *Nature*, 420(6912), 171–173.

- <https://doi.org/10.1038/nature01030>
- Dell'Aglio, D. D., Mena, S., Mauxion, R., McMillan, W. O., & Montgomery, S. H. (2022). Divergence in *Heliconius* flight behaviour is associated with local adaptation to different forest structures. *Journal of Animal Ecology*, *91*(4), 727–737. <https://doi.org/10.1111/1365-2656.13675>
- Edomwande, C., & Barbosa, F. (2020). The influence of predation risk on mate signaling and mate choice in the lesser waxmoth *Achroia grisella*. *Scientific Reports*, *10*(1), 524. <https://doi.org/10.1038/s41598-020-57481-1>
- Engler-Chauat, H. S., & Gilbert, L. E. (2007). De novo Synthesis vs. Sequestration: Negatively correlated metabolic traits and the evolution of host plant specialization in cyanogenic butterflies. *Journal of Chemical Ecology*, *33*(1), 25–42. <https://doi.org/10.1007/s10886-006-9207-8>
- Faure, P. A., & Hoy, R. R. (2000). The sounds of silence: Cessation of singing and song pausing are ultrasound-induced acoustic startle behaviors in the katydid *Neoconocephalus ensiger* (Orthoptera; Tettigoniidae). *Journal of Comparative Physiology A: Sensory, Neural, and Behavioral Physiology*, *186*(2), 129–142. <https://doi.org/10.1007/s003590050013>
- Finkbeiner, S. D., Briscoe, A. D., & Reed, R. D. (2012). The benefit of being a social butterfly: Communal roosting deters predation. *Proceedings of the Royal Society B: Biological Sciences*, *279*(1739), 2769–2776. <https://doi.org/10.1098/rspb.2012.0203>
- Fisher, K. A., & Stankowich, T. (2018). Antipredator strategies of striped skunks in response to cues of aerial and terrestrial predators. *Animal Behaviour*, *143*, 25–34. <https://doi.org/10.1016/j.anbehav.2018.06.023>

- Fitzpatrick, J. W. (1980). Foraging behavior of neotropical tyrant flycatchers. *The Condor*, 82(1), 43–57. <https://doi.org/10.2307/1366784>
- Fournier, J. P., Dawson, J. W., Mikhail, A., & Yack, J. E. (2013). If a bird flies in the forest, does an insect hear it? *Biology Letters*, 9(5), 20130319. <https://doi.org/10.1098/rsbl.2013.0319>
- Gall, L. F. (1984). The effects of capturing and marking on subsequent activity in *Boloria acrocnema* (Lepidoptera: Nymphalidae), with a comparison of different numerical models that estimate population size. *Biological Conservation*, 28(2), 139–154. [https://doi.org/10.1016/0006-3207\(84\)90032-6](https://doi.org/10.1016/0006-3207(84)90032-6)
- Greene, E., & Meagher, T. (1998). Red squirrels, *Tamiasciurus hudsonicus*, produce predator-class specific alarm calls. *Animal Behaviour*, 55(3), 511–518. <https://doi.org/10.1006/anbe.1997.0620>
- Hines, H. M., Counterman, B. A., Papa, R., Albuquerque De Moura, P., Cardoso, M. Z., Linares, M., Mallet, J., Reed, R. D., Jiggins, C. D., Kronforst, M. R., & McMillan, W. O. (2011). Wing patterning gene redefines the mimetic history of *Heliconius* butterflies. *Proceedings of the National Academy of Sciences*, 108(49), 19666–19671. <https://doi.org/10.1073/pnas.1110096108>
- Jacobs, D. S., Ratcliffe, J. M., & Fullard, J. H. (2008). Beware of bats, beware of birds: The auditory responses of eared moths to bat and bird predation. *Behavioral Ecology*, 19(6), 1333–1342. <https://doi.org/10.1093/beheco/arn071>
- Klein, A. L., & De Araújo, A. M. (2010). Courtship behavior of *Heliconius erato phyllis* (Lepidoptera, Nymphalidae) towards virgin and mated females: Conflict between attraction and repulsion signals? *Journal of Ethology*, 28(3), 409–420. <https://doi.org/10.1007/s10164-010-0209-1>



- Lane, K. A., Lucas, K. M., & Yack, J. E. (2008). Hearing in a diurnal, mute butterfly, *Morpho peleides* (Papilionoidea, Nymphalidae). *The Journal of Comparative Neurology*, *508*(5), 677–686. <https://doi.org/10.1002/cne.21675>
- Langham, G. M. (2004). Specialized avian predators repeatedly attack novel color morphs of *Heliconius* butterflies. *Evolution*, *58*(12), 2783–2787.
- Langham, G. M. (2006). Rufous-tailed jacamars and aposematic butterflies: Do older birds attack novel prey? *Behavioral Ecology*, *17*(2), 285–290.  
<https://doi.org/10.1093/beheco/ari027>
- Lea, A. J., & Blumstein, D. T. (2011). Age and sex influence marmot antipredator behavior during periods of heightened risk. *Behavioral Ecology and Sociobiology*, *65*(8), 1525–1533. <https://doi.org/10.1007/s00265-011-1162-x>
- Lind, J., & Cresswell, W. (2005). Determining the fitness consequences of antipredation behavior. *Behavioral Ecology*, *16*(5), 945–956.  
<https://doi.org/10.1093/beheco/ari075>
- Lohrey, A. K., Clark, D. L., Gordon, S. D., & Uetz, G. W. (2009). Antipredator responses of wolf spiders (Araneae: Lycosidae) to sensory cues representing an avian predator. *Animal Behaviour*, *77*(4), 813–821.  
<https://doi.org/10.1016/j.anbehav.2008.12.025>
- Lucas, K. M., Mongrain, J. K., Windmill, J. F. C., Robert, D., & Yack, J. E. (2014). Hearing in the crepuscular owl butterfly (*Caligo eurilochus*, Nymphalidae). *Journal of Comparative Physiology A*, *200*(10), 891–898.  
<https://doi.org/10.1007/s00359-014-0933-z>
- Lucas, K. M., Windmill, J. F. C., Robert, D., & Yack, J. E. (2009). Auditory mechanics and sensitivity in the tropical butterfly *Morpho peleides* (Papilionoidea,

- Nymphalidae). *Journal of Experimental Biology*, 212(21), 3533–3541.  
<https://doi.org/10.1242/jeb.032425>
- Mallet, J., & Gilbert, L. E. (1995). Why are there so many mimicry rings? Correlations between habitat, behaviour and mimicry in *Heliconius* butterflies. *Biological Journal of the Linnean Society*, 55, 159–180.
- Mikhail, A., Lewis, J. E., & Yack, J. E. (2018). What does a butterfly hear? Physiological characterization of auditory afferents in *Morpho peleides* (Nymphalidae). *Journal of Comparative Physiology A*, 204(9–10), 791–799.  
<https://doi.org/10.1007/s00359-018-1280-2>
- Morton, E. S. (1971). Food and migration habits of the Eastern kingbird in Panama. *The Auk*, 88(4), 925–926. <https://doi.org/10.2307/4083855>
- Olofsson, M., Eriksson, S., Jakobsson, S., & Wiklund, C. (2012). Deimatic display in the European swallowtail butterfly as a secondary defence against attacks from Great tits. *PLoS ONE*, 7(10), e47092. <https://doi.org/10.1371/journal.pone.0047092>
- Olofsson, M., Vallin, A., Jakobsson, S., & Wiklund, C. (2011). Winter predation on two species of hibernating butterflies: Monitoring rodent attacks with infrared cameras. *Animal Behaviour*, 81(3), 529–534.  
<https://doi.org/10.1016/j.anbehav.2010.12.012>
- Palmer, M. S., & Packer, C. (2021). Reactive anti-predator behavioral strategy shaped by predator characteristics. *PLOS ONE*, 16(8), e0256147.  
<https://doi.org/10.1371/journal.pone.0256147>
- Pinheiro, C. E. G. (1996). Palatability and escaping ability in neotropical butterflies: Tests with wild kingbirds ( *Tyrannus melancholicus*, Tyrannidae). *Biological*

- Journal of the Linnean Society*, 59(4), 351–365. <https://doi.org/10.1111/j.1095-8312.1996.tb01471.x>
- Pinheiro, C. E. G. (2011). On the evolution of warning coloration, Batesian and Müllerian mimicry in Neotropical butterflies: The role of jacamars (Galbulidae) and tyrant-flycatchers (Tyrannidae). *Journal of Avian Biology*, 42(4), 277–281. <https://doi.org/10.1111/j.1600-048X.2011.05435.x>
- Pinheiro, C. E. G., & Campos, V. C. (2019). The responses of wild jacamars (*Galbula ruficauda*, Galbulidae) to aposematic, aposematic and cryptic, and cryptic butterflies in central Brazil. *Ecological Entomology*, 44(4), 441–450. <https://doi.org/10.1111/een.12723>
- Pinheiro, C. E. G., & Cintra, R. (2017). Butterfly predators in the neotropics: Which birds are involved? *Journal of the Lepidopterists' Society*, 71(2), 109–114. <https://doi.org/10.18473/lepi.71i2.a5>
- Pinheiro De Castro, É. C., Zagrobelny, M., Zurano, J. P., Zikan Cardoso, M., Feyereisen, R., & Bak, S. (2019). Sequestration and biosynthesis of cyanogenic glucosides in passion vine butterflies and consequences for the diversification of their host plants. *Ecology and Evolution*, 9, 5079–5093. <https://doi.org/10.1002/ece3.5062>
- Prakash, H., Greif, S., Yovel, Y., & Balakrishnan, R. (2021). Acoustically eavesdropping bat predators take longer to capture katydid prey signalling in aggregation. *Journal of Experimental Biology*, 224(10), jeb233262. <https://doi.org/10.1242/jeb.233262>
- Rather, P. A., Herzog, A. E., Ernst, D. A., & Westerman, E. L. (2022). Effect of experience on mating behaviour in male *Heliconius melpomene* butterflies. *Animal Behaviour*, 183, 139–149. <https://doi.org/10.1016/j.anbehav.2021.11.004>

- Ribarič, D., & Gogala, M. (1996). Acoustic behaviour of some butterfly species of the Genus *Erebia* (Lepidoptera: Satyridae). *Acta Entomologica Slovenica*, 4(1), 5–12.
- Robertson, D. N., Sullivan, T. J., & Westerman, E. L. (2020). Lack of sibling avoidance during mate selection in the butterfly *Bicyclus anynana*. *Behavioural Processes*, 173, 104062. <https://doi.org/10.1016/j.beproc.2020.104062>
- Rojas, B., Mappes, J., & Burdfield-Steel, E. (2019). Multiple modalities in insect warning displays have additive effects against wild avian predators. *Behavioral Ecology and Sociobiology*, 73(3), 37. <https://doi.org/10.1007/s00265-019-2643-6>
- Rosen, M. J., Levin, E. C., & Hoy, R. R. (2009). The cost of assuming the life history of a host: Acoustic startle in the parasitoid fly *Ormia ochracea*. *Journal of Experimental Biology*, 212(24), 4056–4064. <https://doi.org/10.1242/jeb.033183>
- Sitvarin, M. I., & Rypstra, A. L. (2012). Sex-Specific response of *Pardosa milvina* (Araneae: Lycosidae) to experience with a chemotactile predation cue. *Ethology*, 118(12), 1230–1239. <https://doi.org/10.1111/eth.12029>
- Srygley, R. B. (1994). Locomotor mimicry in butterflies? The associations of positions of Centres of Mass among Groups of Mimetic, Unprofitable Prey. *Philosophical Transactions: Biological Sciences*, 343(1304), 145–155.
- Sun, P., Mhatre, N., Mason, A. C., & Yack, J. E. (2018). In that vein: Inflated wing veins contribute to butterfly hearing. *Biology Letters*, 14(10), 20180496. <https://doi.org/10.1098/rsbl.2018.0496>
- Swihart, S. L. (1967). Hearing in Butterflies (Nymphalidae: *Heliconius*, *Ageronia*). *Journal of Insect Physiology*, 13, 469–476.
- Thaker, M., Lima, S. L., & Hews, D. K. (2009). Alternative antipredator tactics in tree lizard morphs: Hormonal and behavioural responses to a predator encounter.

- Animal Behaviour*, 77(2), 395–401. <https://doi.org/10.1016/j.anbehav.2008.10.014>
- Torsekar, V. R., Isvaran, K., & Balakrishnan, R. (2019). Is the predation risk of mate-searching different between the sexes? *Evolutionary Ecology*, 33(3), 329–343. <https://doi.org/10.1007/s10682-019-09982-3>
- Tribblehorn, J. D., Ghose, K., Bohn, K., Moss, C. F., & Yager, D. D. (2008). Free-flight encounters between praying mantids (*Parasphendale agrionina*) and bats (*Eptesicus fuscus*). *Journal of Experimental Biology*, 211(4), 555–562. <https://doi.org/10.1242/jeb.005736>
- Umbers, K. D. L., & Mappes, J. (2015). Postattack deimatic display in the mountain katydid, *Acripeza reticulata*. *Animal Behaviour*, 100, 68–73. <https://doi.org/10.1016/j.anbehav.2014.11.009>
- Vallin, A., Jakobsson, S., Lind, J., & Wiklund, C. (2006). Crypsis versus intimidation—Anti-predation defence in three closely related butterflies. *Behavioral Ecology and Sociobiology*, 59(3), 455–459. <https://doi.org/10.1007/s00265-005-0069-9>
- Westerman, E. L., Chirathivat, N., Schyling, E., & Monteiro, A. (2014). Mate preference for a phenotypically plastic trait is learned, and may facilitate preference-phenotype matching. *Evolution*, 68(6), 1661–1670. <https://doi.org/10.1111/evo.12381>
- Wickham, H. (2016). *ggplot2: Elegant Graphics for Data Analysis*. Springer-Verlag New York. <https://ggplot2.tidyverse.org>
- Wormington, J. D., & Juliano, S. A. (2014). Hunger-dependent and sex-specific antipredator behaviour of larvae of a size-dimorphic mosquito. *Ecological Entomology*, 39(5), 548–555. <https://doi.org/10.1111/een.12129>

Zaguri, M., & Hawlena, D. (2020). Odours of non-predatory species help prey moderate their risk assessment. *Functional Ecology*, 34(4), 830–839.

<https://doi.org/10.1111/1365-2435.13509>

590

591

592

593

594

595

596

597

598

599

600

601

602

603

604

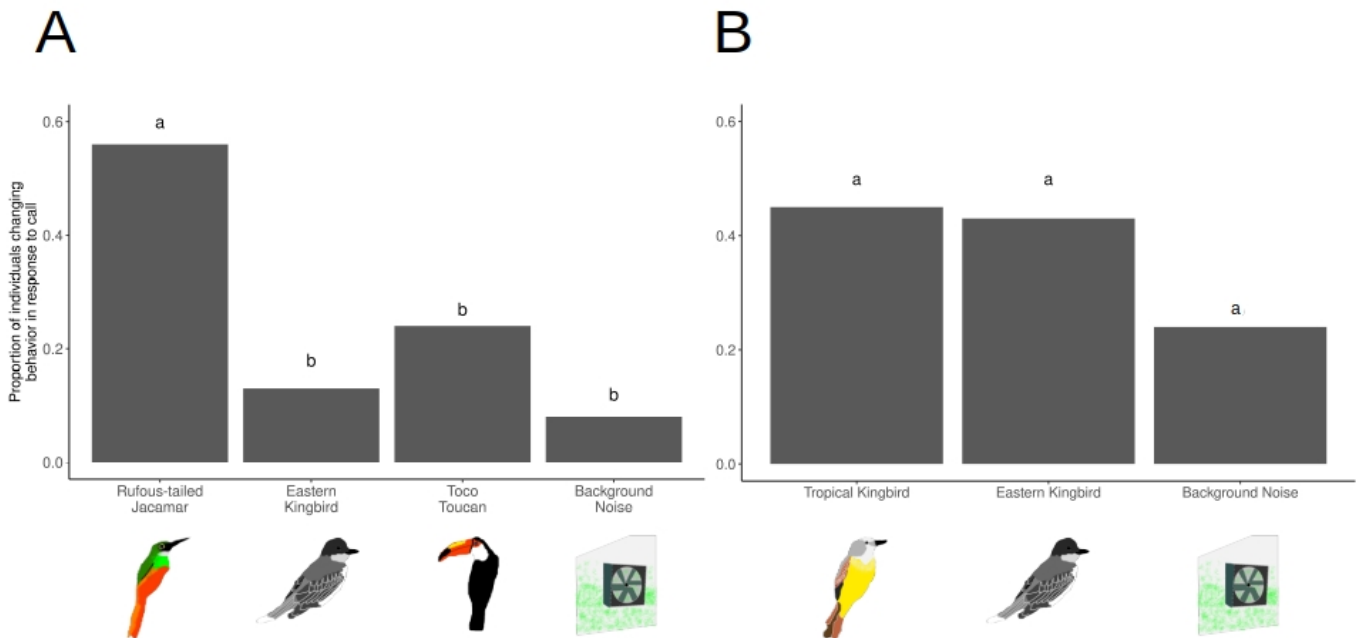
605

606

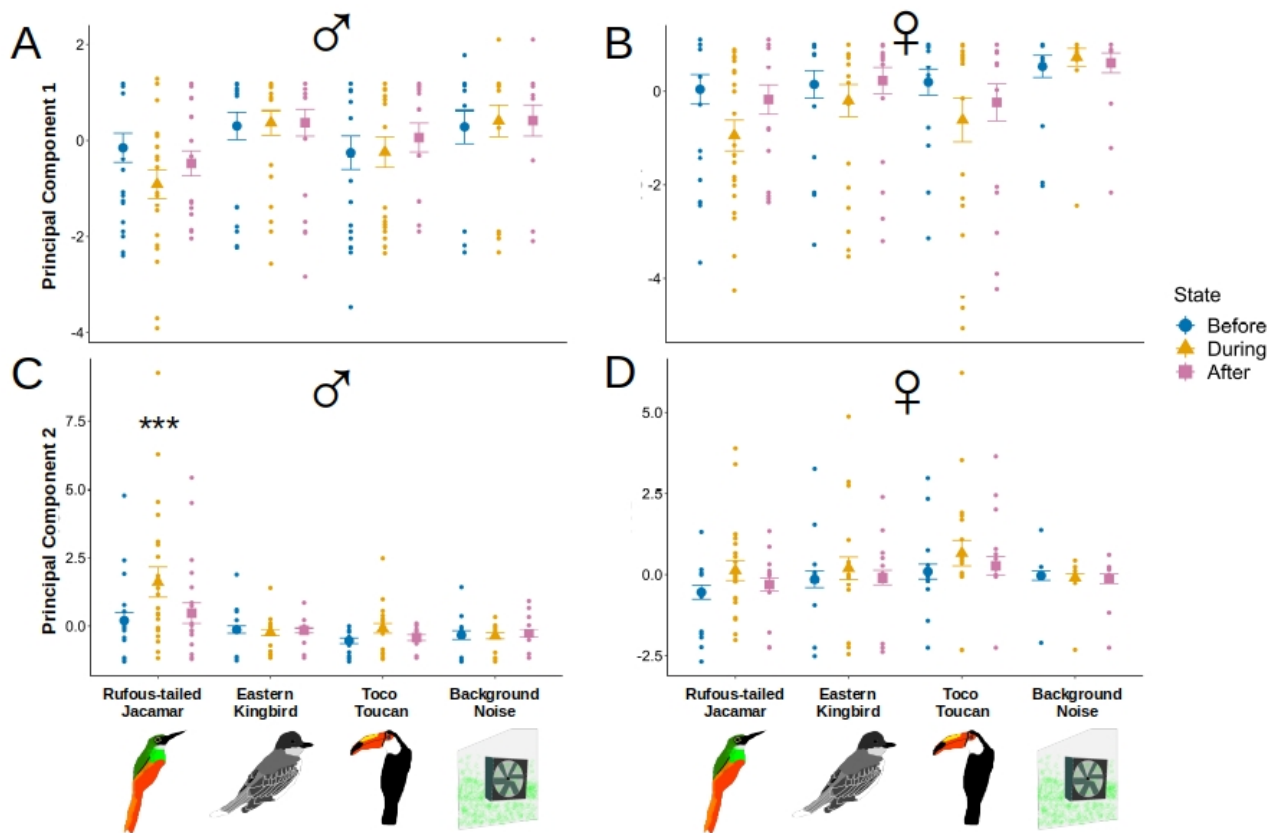
607

608

609 **Figures:**

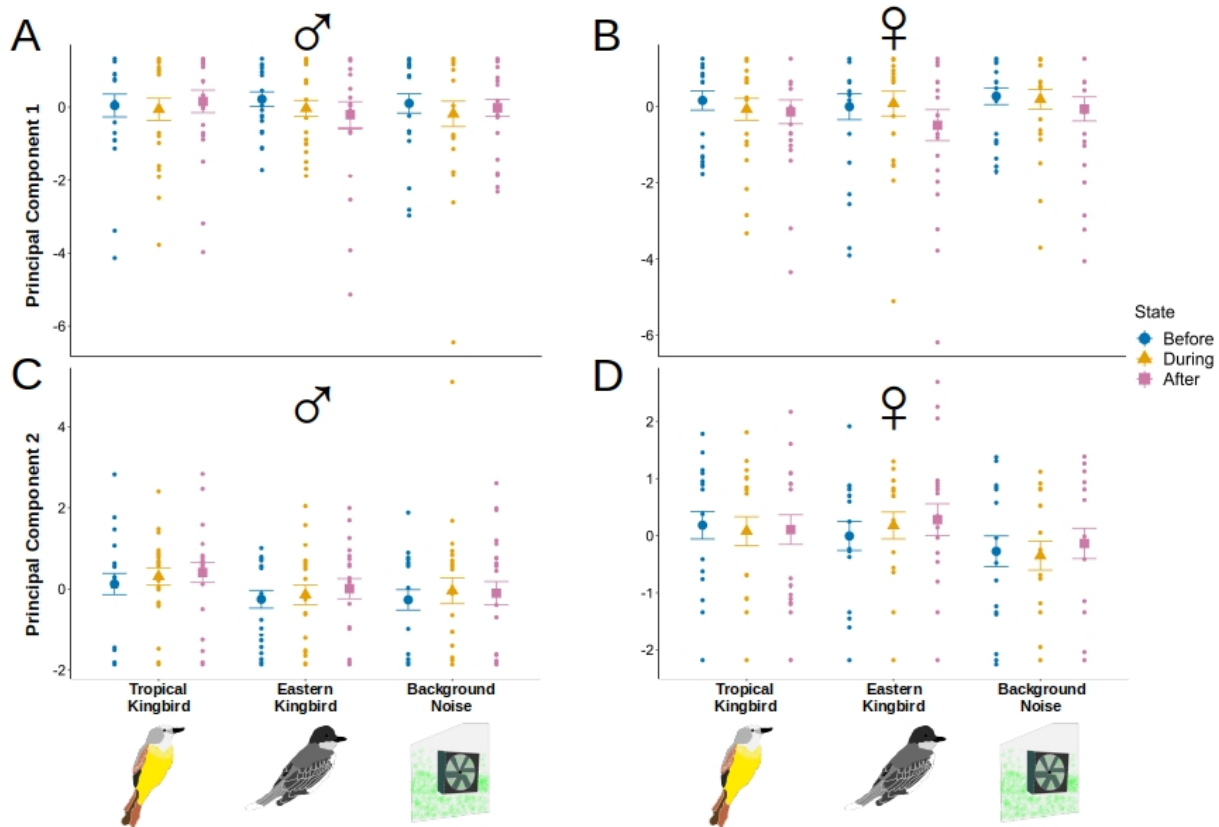


610 **Figure 1:** Proportion of *H. m. plessini* individuals changing behaviour in response to  
611 calls (between before start and after end of calls) for A) experiment 1; B) experiment 2;  
612 Different letters on each bars indicate statistical significance at  $p < 0.05$ .



613 **Figure 2:** Mean  $\pm$  SE of principal component variables for male and female *H. m.*  
614 *plessini* for a minute before, during and after calls. A) PC 1 in males for experiment 1; B)  
615 PC 1 in females for experiment 1; C) PC 2 in males for experiment 1; D) PC 2 in females  
616 for experiment 1. \*\*\* indicates significance with  $p < 0.0001$ .





617 **Figure 3:** Mean  $\pm$  SE of principal component variables for male and female *H. m.*  
618 *plessini* for a minute before, during and after calls. A) PC 1 in males for experiment 2; B)  
619 PC 1 in females for experiment 2; C) PC 2 in males for experiment 2; D) PC 2 in females  
620 for experiment 2. None of them are significantly different from each other.

621 **Tables:**

622

623 **Table 1: GLMM results on the effect of treatment (calls) and sex on proportion of**  
624 **butterflies changing their behaviour in response to calls. p<0.05 are bolded**

625

626

Fixed effect	Estimate	SE	z value	Pr (> z )
<b>Intercept</b>	<b>-2.55</b>	<b>0.64</b>	<b>-3.98</b>	<b>&lt;0.0001</b>
<b>Treatment (Rufous-tailed Jacamar)</b>	<b>2.67</b>	<b>0.67</b>	<b>3.96</b>	<b>&lt;0.0001</b>
Treatment (Eastern Kingbird)	0.5	0.74	0.67	0.640
Treatment (Toco Toucan)	1.24	0.69	1.78	0.777
Sex (male)	0.29	0.38	0.76	0.44
Random effect				
Order (Intercept)	1.7e-15	4.1e-8		

627

628

629

630

631

632

633

634

635

636

637

638

639

640

641

642

643

644

645

646

647

648

649

650 **Table 2: Pairwise differences in the proportion of individuals changing their**  
651 **behavioural state in response to calls in experiment 1. p<0.05 are bolded**  
652

---

ANOVA Type II Wald Chisquare test

**Treatment:  $\chi^2 = 27.12$ ; df = 3; p-value < 0.0001**

Sex:  $\chi^2 = 0.577$ ; df = 1; p-value = 0.44

---

Pairwise comparisons between treatment

---

Group 1	Group 2	p-value	Adj. p-value
<b>Rufous-tailed Jacamar</b>	<b>Eastern Kingbird</b>	<b>0.00002</b>	<b>0.0001</b>
<b>Rufous-tailed Jacamar</b>	<b>Toucan</b>	<b>0.002</b>	<b>0.01</b>
<b>Rufous-tailed Jacamar</b>	<b>Greenhouse noise</b>	<b>0.000005</b>	<b>0.00003</b>
Eastern Kingbird	Toco Toucan	0.28	1
Eastern Kingbird	Greenhouse noise	0.72	1
Toco Toucan	Greenhouse noise	0.07	0.47

---

653  
654  
655  
656  
657  
658

659 **Table 3: Effect of treatment (rufous-tailed jacamar, Eastern kingbird, toco toucan**  
 660 **and greenhouse background noise calls), state (one minute before, during and after**  
 661 **call) and their interaction on male and female PC1 and PC2 in experiment 1. p<0.05**  
 662 **are bolded**  
 663

♂	AIC	Df	F value	Pr (>F)	♀	AIC	Df	F value	Pr (>F)
PC1	884				PC1	910			
<b>Treatment</b>		<b>3</b>	<b>5.887</b>	<b>0.0006</b>	<b>Treatment</b>		<b>3</b>	<b>5.090</b>	<b>0.001</b>
State		2	0.447	0.640	State		2	2.794	0.063
Treatment*State		6	0.540	0.777	Treatment*State		6	0.732	0.624
PC2	781				PC2	805			
<b>Treatment</b>		<b>3</b>	<b>13.900</b>	<b>2.2e-08</b>	<b>Treatment</b>		<b>3</b>	<b>2.759</b>	<b>0.043</b>
<b>State</b>		<b>2</b>	<b>3.649</b>	<b>0.0275</b>	State		2	2.498	0.084
<b>Treatment*State</b>		<b>6</b>	<b>2.336</b>	<b>0.0328</b>	Treatment*State		6	0.373	0.896
PC3	787				PC3	745			
Treatment		3	1.432	0.234	Treatment		3	2.017	0.112
State		2	0.856	0.426	State		2	1.071	0.344
Treatment*State		6	0.697	0.652	Treatment*State		6	0.577	0.748
Courtship	239				Copulation	1692			
Treatment		3	0.984	0.401	<b>Treatment</b>		<b>3</b>	<b>2.765</b>	<b>0.042</b>
State		2	0.999	0.370	State		2	0	1
Treatment*State		6	0.984	0.437	Treatment*State		6	0	1
Sitting near other	1659				Abdomen lift				
<b>Treatment</b>		<b>3</b>	<b>3.500</b>	<b>0.016</b>	Treatment		3	0.922	0.431
State		2	0.011	0.988	State		2	0.997	0.370
Treatment*State		6	0.003	1	Treatment*State		6	0.922	0.480

664  
 665

666

## Supplementary Material 1

667

### Behavioural changes in aposematic *Heliconius melpomene* butterflies in response to

668

### their predatory bird calls

669

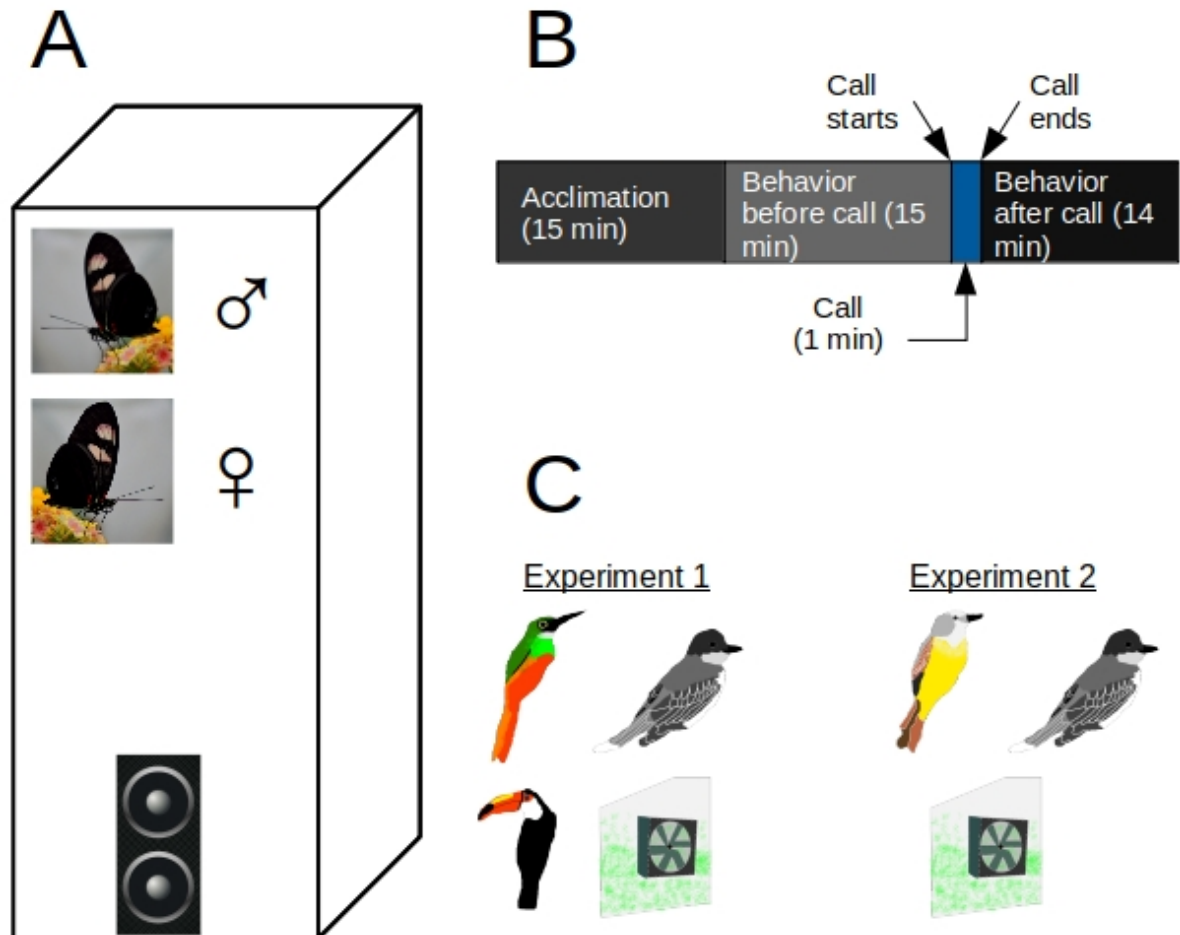
#### Table of Content

670

Name	Brief description	Pg.no
Supplementary Figure 1	Experimental design	3
Supplementary Figure 2	Spectograms of calls	4
Supplementary Figure 3	Proportion changing behaviour at start of call	6
Supplementary Figure 4	Proportion changing behaviour at end of call	7
Supplementary Figure 5	Long term effect of calls on PC1 and PC2 behavior in expt 1	8
Supplementary Figure 6	Long term effect of calls on PC1 and PC2 behavior in expt 2	9
Supplementary Table 1	GLMM results for behavioural change at start of call in expt 1	10
Supplementary Table 2	Pairwise difference in behavioural change at start of call in expt 1	11
Supplementary Table 3	GLMM results for behavioural change at end of call in expt 1	12
Supplementary Table 4	Pairwise difference in behavioural change at end of call in expt 1	13
Supplementary Table 5	PC loadings for males for 3 minutes in expt 1	14
Supplementary Table 6	PC loadings for females for 3 minutes in expt 1	15
Supplementary Table 7	ANOVA post-hoc test for 3 minutes in expt 1	16
Supplementary Table 8	PC loadings for males for 28 minutes in expt 1	17
Supplementary Table 9	PC loadings for females for 28 minutes in expt 1	18
Supplementary Table 10	ANOVA models for PC1, PC2, PC3, and inter-sexual behaviours for 28 minutes in expt 1	19
Supplementary Table 11	ANOVA post-hoc test for 28 minutes in expt 1	20
Supplementary Table 12	GLMM results for behavioural change at start of call in expt 2	21
Supplementary Table 13	Pairwise difference in behavioural change at start of call in expt 2	22
Supplementary Table 14	GLMM results for behavioural change at end of call in expt 2	23
Supplementary Table 15	Pairwise difference in behavioural change at end of call in expt 2	24
Supplementary Table 16	GLMM results for behavioural change in response to call in expt 2	25

Supplementary Table 17	Pairwise difference in behavioural change in response to call in expt 2	26
Supplementary Table 18	PC loadings for males for 3 minutes in expt 2	27
Supplementary Table 19	PC loadings for females for 3 minutes in expt 2	28
Supplementary Table 20	ANOVA models for PC1, PC2, PC3, and inter-sexual behaviours for 3 minutes in expt 2	29
Supplementary Table 21	ANOVA post-hoc test for 3 minutes in expt 2	30
Supplementary Table 22	PC loadings for males for 28 minutes in expt 2	31
Supplementary Table 23	PC loadings for females for 28 minutes in expt 2	32
Supplementary Table 24	ANOVA models for PC1, PC2, PC3, and inter-sexual behaviours for 28 minutes in expt 2	33
Supplementary Table 25	ANOVA post-hoc test for 28 minutes in expt 2	34

671  
672  
673  
674  
675  
676  
677

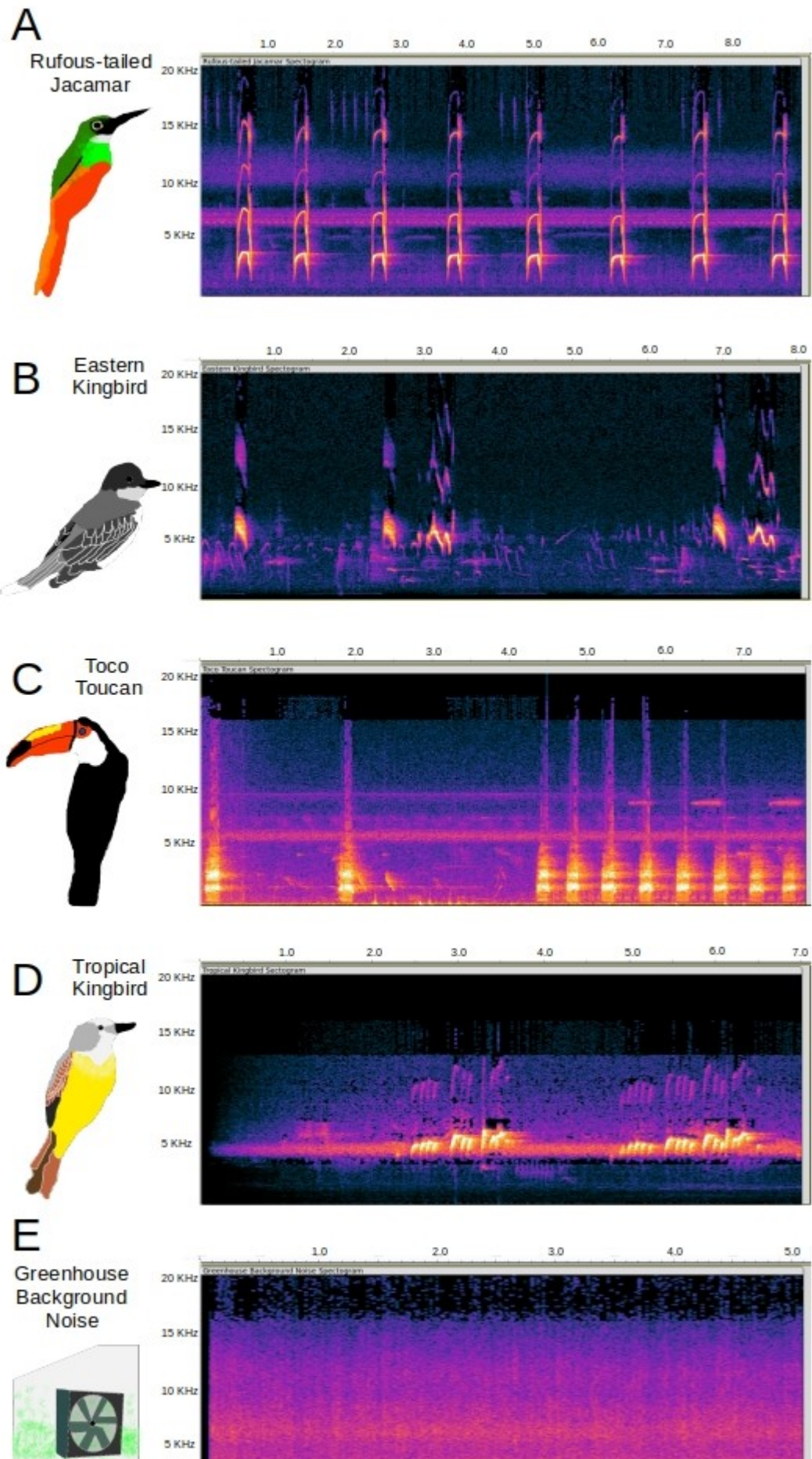


**Supplementary Figure 1:** Experimental design. A) 3-15-day male and female *H. m. plessini* butterflies were subjected in an experimental cage with a blue tooth speaker and a *Lantana spp.* plant during each experimental assay. B) The timeline of each assay conducted where the butterflies were acclimated for 15 minutes and their behaviours recorded for the next 30 minutes. During the 16<sup>th</sup> minute, a call was randomly played for a minute. C) The calls used in the two experiments in this study. Clockwise from top left in experiment 1: rufous-tailed jacamar, Eastern kingbird, greenhouse background noise, and toco toucan. Clockwise from top left in experiment 2: tropical kingbird, Eastern kingbird, and greenhouse background noise.

678

679

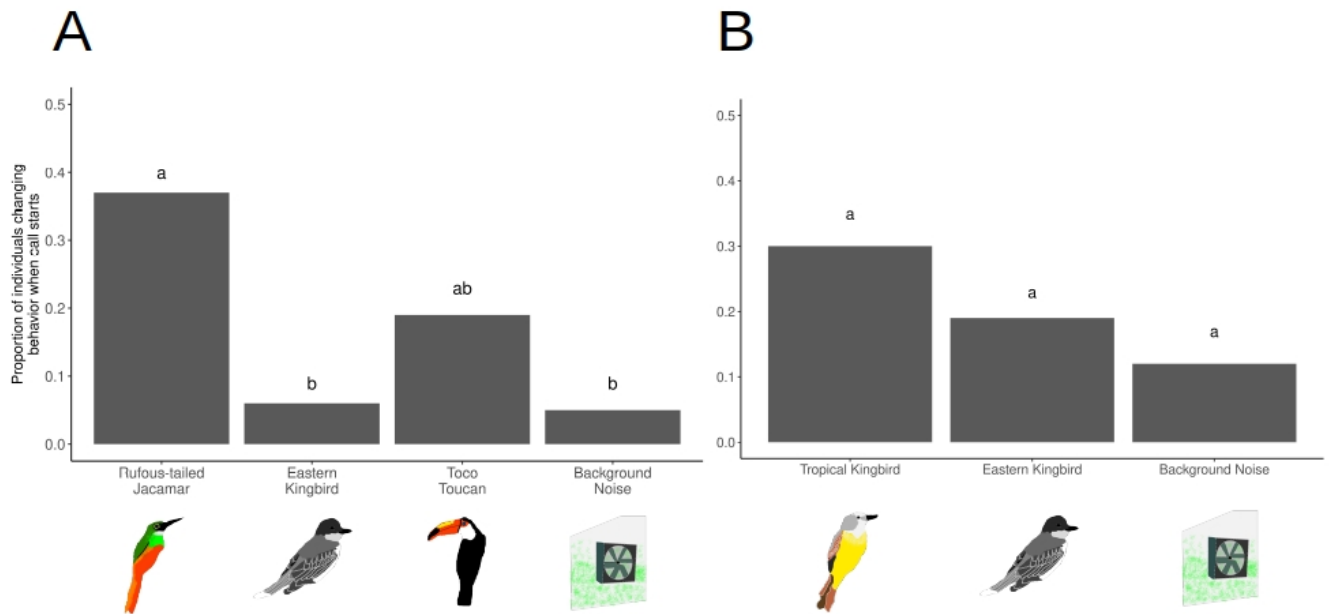






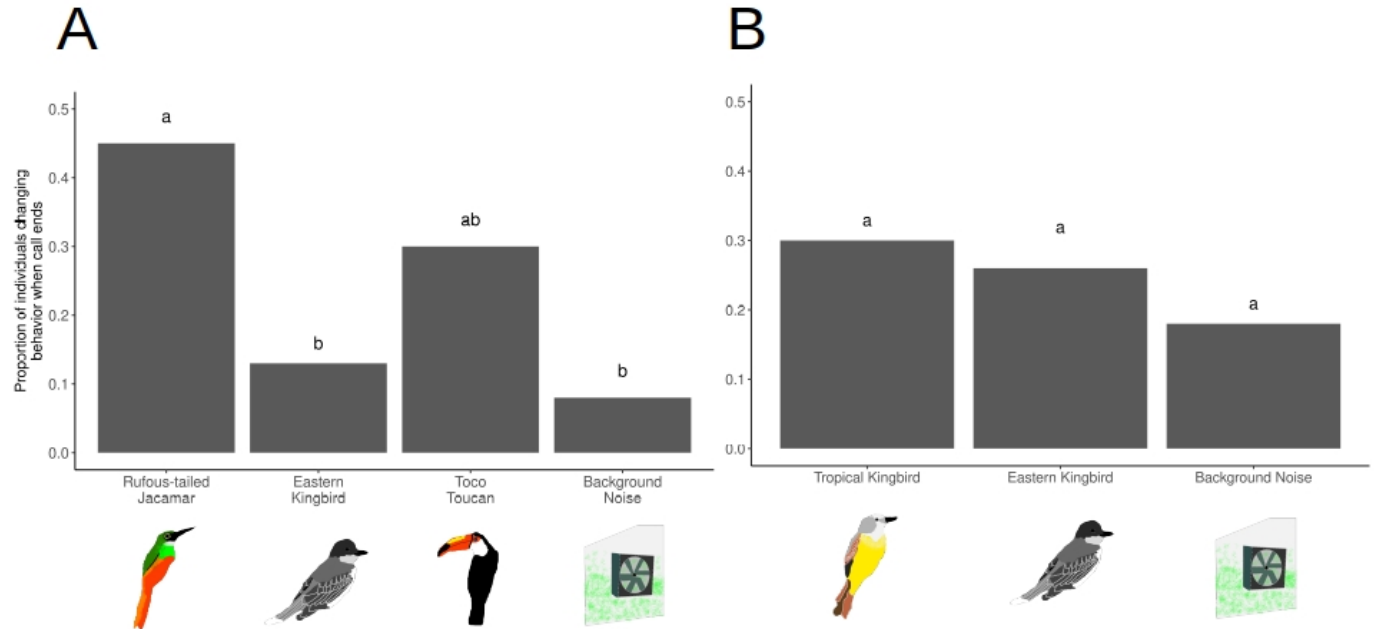
**Supplementary Figure 2:** Spectrograms of the calls used during this study A) rufous-tailed jacamar; B) Eastern kingbird; C) toco toucan; D) tropical kingbird; E) greenhouse background noise.

680  
681  
682  
683  
684  
685  
686  
687  
688  
689  
690  
691  
692  
693  
694  
695  
696  
697  
698  
699  
700  
701  
702  
703  
704  
705  
706  
707  
708  
709  
710  
711  
712  
713  
714  
715  
716  
717  
718



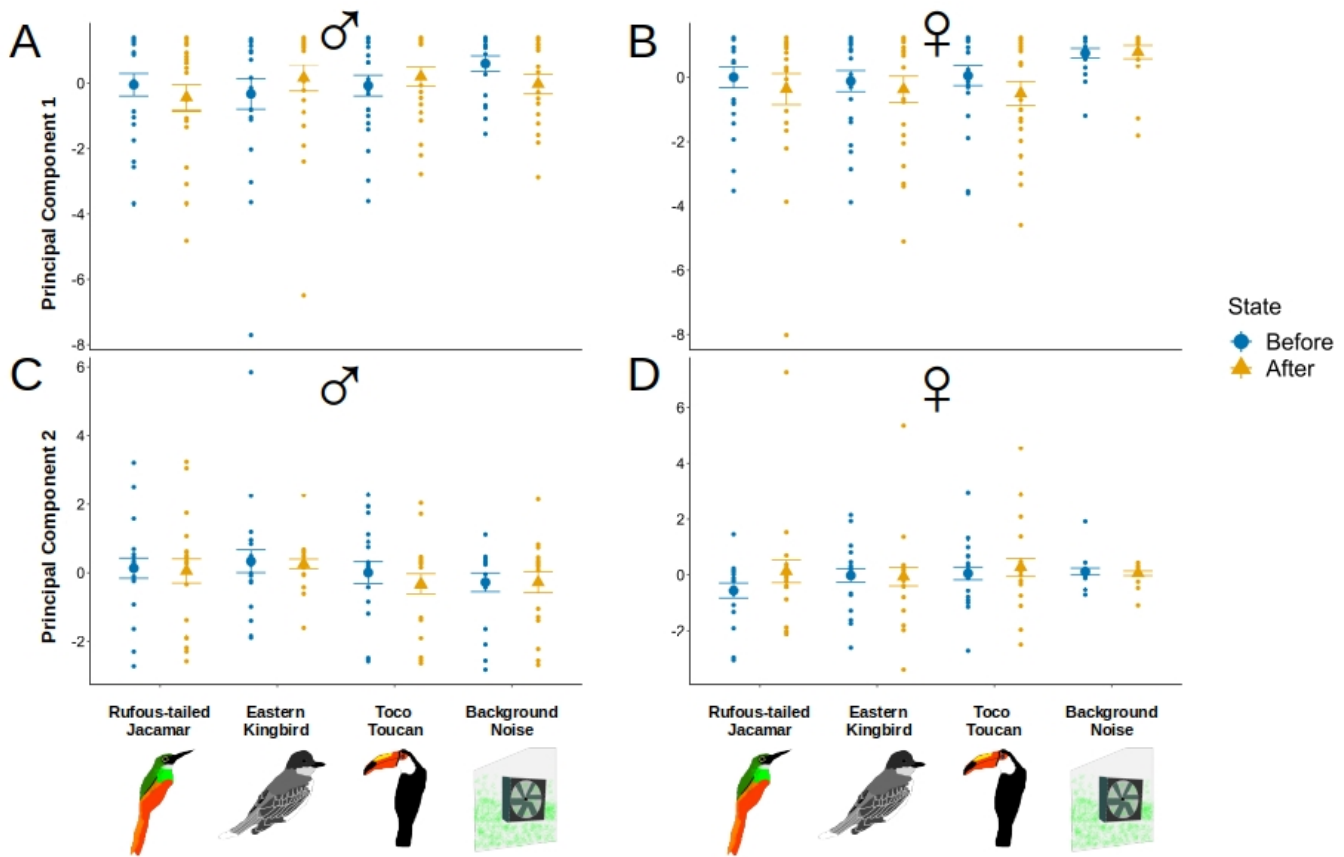
**Supplementary Figure 3:** Proportion of *H. m. plessini* individuals changing behaviour in response to the start of the calls (between before start and after start of calls) for A) experiment 1; B) experiment 2; Different letters on each bars indicate statistical significance at  $p < 0.05$ .

719  
720  
721  
722  
  
723  
724  
725  
726  
727  
728  
729  
730  
731  
732



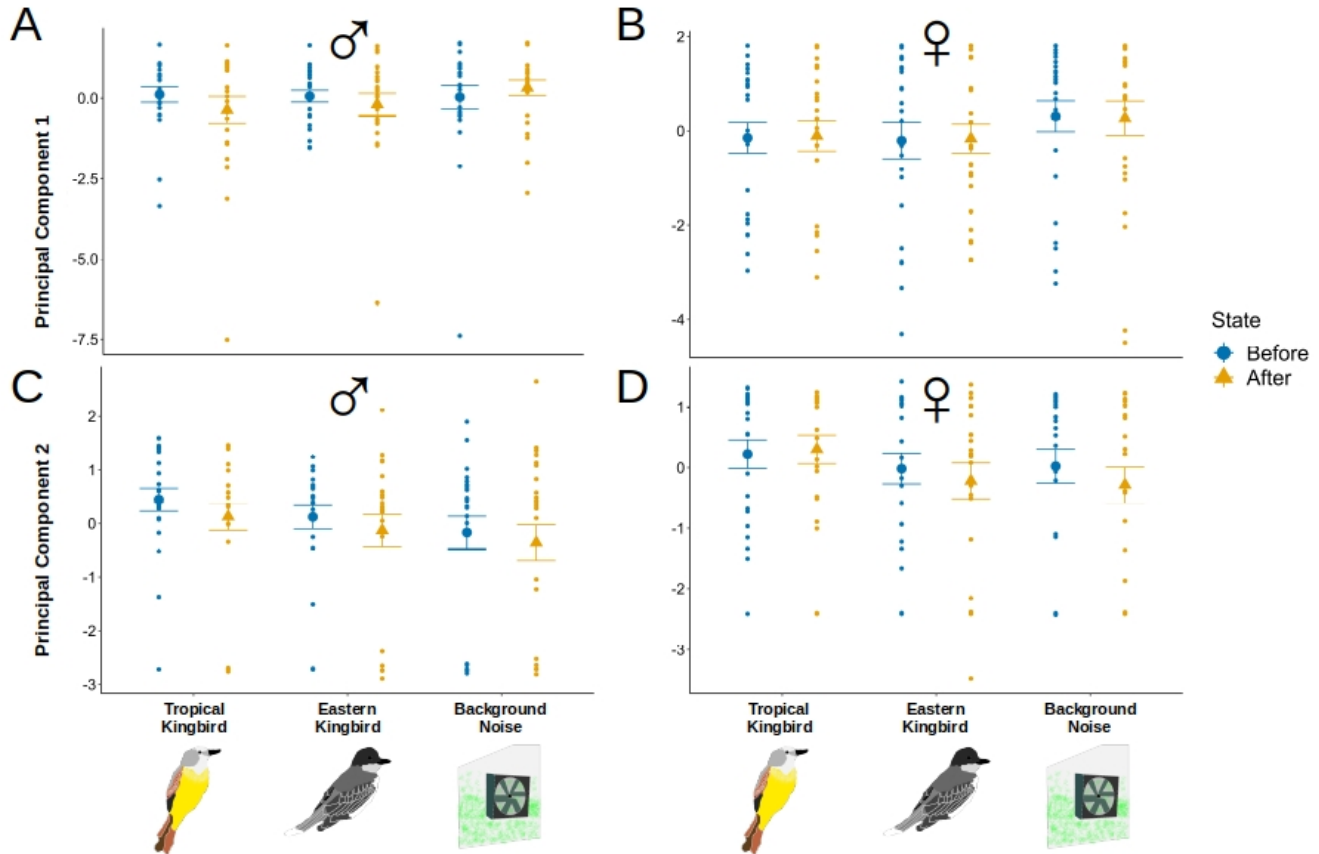
**Supplementary Figure 4:** Proportion of *H. m. plessini* individuals changing behaviour in response to the end of the calls (between before end and after end of calls) for A) experiment 1; B) experiment 2; Different letters on each bars indicate statistical significance at  $p < 0.05$ .

733  
734  
735  
736  
737  
738  
739  
740  
741  
742  
743  
744  
745  
746



**Supplementary Figure 5:** Mean  $\pm$  SE of principal component variables for male and female *H. m. plessini* for 14 minutes before, and after calls. A) PC 1 in males for experiment 1; B) PC 1 in females for experiment 1; C) PC 2 in males for experiment 1; D) PC 2 in females for experiment 1. None of them are significantly different from each other.

747  
748  
749  
750  
751  
752  
753  
754  
755  
756  
757  
758



**Supplementary Figure 6:** Mean  $\pm$  SE of principal component variables for male and female *H. m. plessini* for 14 minutes before, and after calls. A) PC 1 in males for experiment 2; B) PC 1 in females for experiment 2; C) PC 2 in males for experiment 2; D) PC 2 in females for experiment 2. None of them are significantly different from each other.

759  
760  
761  
762  
763  
764  
765  
766  
767  
768  
769  
770

771 **Supplementary Table 1: GLMM results on the effect of treatment (calls) and sex on**  
772 **proportion of butterflies changing their behaviour at the start of calls in experiment**  
773 **1. p<0.05 are bolded**

774  
775

Fixed effect	Estimate	SE	z value	Pr (> z )
<b>Intercept</b>	<b>-2.97</b>	<b>0.76</b>	<b>-3.88</b>	<b>&lt;0.001</b>
<b>Treatment (Rufous-tailed Jacamar)</b>	<b>2.30</b>	<b>0.78</b>	<b>2.91</b>	<b>&lt;0.001</b>
Treatment (Eastern Kingbird)	0.17	0.94	0.18	0.856
Treatment (Toco Toucan)	1.42	0.81	1.73	0.082
Sex (male)	0.26	0.42	0.62	0.529
Random effect				
Order (Intercept)	0	0		

776  
777  
778  
779  
780  
781  
782  
783  
784  
785  
786  
787  
788  
789  
790  
791  
792  
793  
794  
795  
796  
797  
798  
799

800 **Supplementary Table 2: Pairwise differences in the proportion of individuals**  
801 **changing their behavioural state in at the start of calls in experiment 1. p<0.05 are**  
802 **bolded**  
803

---

**ANOVA Type II Wald Chisquare test**

**Treatment:  $\chi^2 = 16.03$ ; df = 3; p-value < 0.01**

Sex:  $\chi^2 = 0.396$ ; df = 1; p-value = 0.529

---

Pairwise comparisons between treatment

---

Group 1	Group 2	p-value	Adj. p-value
<b>Rufous-tailed Jacamar</b>	<b>Eastern Kingbird</b>	<b>0.0007</b>	<b>0.004</b>
Rufous-tailed Jacamar	Toucan	0.104	0.624
<b>Rufous-tailed Jacamar</b>	<b>Greenhouse noise</b>	<b>0.001</b>	<b>0.006</b>
Eastern Kingbird	Toco Toucan	0.11	0.7
Eastern Kingbird	Greenhouse noise	1	1
Toco Toucan	Greenhouse noise	0.10	0.61

---

804  
805  
806  
807  
808  
809

810 **Supplementary Table 3: GLMM results on the effect of treatment (calls) and sex on**  
811 **proportion of butterflies changing their behaviour at the end of calls in experiment**  
812 **1. p<0.05 are bolded**

813  
814

Fixed effect	Estimate	SE	z value	Pr (> z )
<b>Intercept</b>	<b>-2.46</b>	<b>0.64</b>	<b>-3.79</b>	<b>&lt;0.001</b>
<b>Treatment (Rufous-tailed Jacamar)</b>	<b>2.22</b>	<b>0.67</b>	<b>3.30</b>	<b>&lt;0.001</b>
Treatment (Eastern Kingbird)	0.44	0.75	0.59	0.554
<b>Treatment (Toco Toucan)</b>	<b>1.62</b>	<b>0.69</b>	<b>2.34</b>	<b>&lt;0.05</b>
Sex (male)	0.13	0.37	0.37	0.709
Random effect				
Order (Intercept)	0.06	0.25		

815  
816  
817  
818  
819  
820  
821  
822  
823  
824  
825  
826  
827  
828  
829  
830  
831  
832  
833  
834  
835  
836  
837  
838



839 **Supplementary Table 4: Pairwise differences in the proportion of individuals**  
840 **changing their behavioural state in at the end of calls in experiment 1. p<0.05 are**  
841 **bolded**  
842

---

ANOVA Type II Wald Chisquare test

Treatment:  $\chi^2 = 17.47$ ; df = 3; p-value < 0.001

Sex:  $\chi^2 = 0.139$ ; df = 1; p-value = 0.709

---

Pairwise comparisons between treatment

---

Group 1	Group 2	p-value	Adj. p-value
<b>Rufous-tailed Jacamar</b>	<b>Eastern Kingbird</b>	<b>0.001</b>	<b>0.006</b>
Rufous-tailed Jacamar	Toucan	0.197	1
<b>Rufous-tailed Jacamar</b>	<b>Greenhouse noise</b>	<b>0.0002</b>	<b>0.001</b>
Eastern Kingbird	Toco Toucan	0.07	0.451
Eastern Kingbird	Greenhouse noise	0.724	1
Toco Toucan	Greenhouse noise	0.015	0.093

---

843  
844  
845  
846  
847  
848

849 **Supplementary Table 5: Loadings of each behaviour in Principal Component (PC)**  
850 **composite variables for males in a minute before, during, and after calls in**  
851 **experiment 1**  
852

Behaviour	PC1	PC2	PC3
Rest	0.673	0.112	0.115
Fly	0.170	0.100	0.689
Bask	0.602	0.294	0.200
Flutter	0.195	0.674	0.088
Court	0.053	0.015	0.600
Copulate	0.095	0.254	0.009
Walk	0.169	0.602	0.140
Antenna wiggle	0.253	0.080	0.281
Sitting near each other	0.108	0.025	0.061
% Variance explained	22.28	17.05	14.43
% Total variance explained	22.28	39.33	53.77

853  
854  
855  
856  
857  
858  
859  
860  
861  
862  
863  
864  
865  
866  
867  
868  
869  
870  
871  
872  
873

874 **Supplementary Table 6: Loadings of each behaviour in Principal Component (PC)**  
875 **composite variables for females in a minute before, during and after calls in**  
876 **experiment 1**

877

878

Behaviour	PC1	PC2	PC3
Rest	0.605	0.269	0.208
Fly	0.193	0.236	0.576
Bask	0.490	0.493	0.136
Flutter	0.362	0.577	0.016
Copulate	0.066	0.162	0.583
Walk	0.355	0.512	0.149
Antenna wiggle	0.304	0.071	0.491
Lifting abdomen	0.028	0.030	0.037
% Variance explained	28.18	17.83	13.77
% Total variance explained	28.18	46.02	59.79

879

880

881

882

883

884

885

886

887

888

889

890

891

892

893

894

895

896

897

898

899

900 **Supplementary Table 7: ANOVA Post-hoc test results with PC as dependent**  
 901 **variables and the treatments (calls), state (before, during, and after) and their**  
 902 **interaction as response variables for males and females in a minute before, during,**  
 903 **and after calls in experiment 1. EK= Eastern kingbird; RJ=Rufous-tailed jacamar;**  
 904 **TT=Toco toucan; GN=Greenhouse background noise; Difference=pairwise**  
 905 **difference; lwr=lower range; upr=upper range; padj= adjusted p-value.**

	♂				♀			
	PC1				PC1			
Treatment	Difference	lwr	upr	padj	Difference	lwr	upr	padj
EK-RJ	<b>0.861</b>	<b>0.235</b>	<b>1.487</b>	<b>0.002</b>	0.418	-0.240	1.077	0.357
TT-RJ	0.368	-0.264	1.001	0.434	0.141	-0.525	0.807	0.947
GN-RJ	<b>0.882</b>	<b>0.223</b>	<b>1.542</b>	<b>0.003</b>	<b>0.982</b>	<b>0.287</b>	<b>1.676</b>	<b>0.001</b>
TT-EK	-0.492	-1.125	0.140	0.185	-0.276	-0.943	0.389	0.705
GN-EK	0.021	-0.638	0.680	0.999	0.564	-0.130	1.258	0.155
GN-TT	0.513	-0.512	1.180	0.192	<b>0.840</b>	<b>0.138</b>	<b>1.542</b>	<b>0.011</b>
State								
During-Before	-0.153	-0.661	0.355	0.757	-0.516	-1.051	0.019	0.061
After-Before	0.040	-0.468	0.548	0.981	-0.129	-0.665	0.406	0.836
After-During	0.193	-0.315	0.701	0.643	0.386	-0.149	0.922	0.206
	PC2				PC2			
Treatment	Difference	lwr	upr	padj	Difference	lwr	upr	padj
EK-RJ	<b>-0.934</b>	<b>-1.443</b>	<b>-0.426</b>	<b>0.00002</b>	0.226	-0.307	0.761	0.690
TT-RJ	<b>-1.108</b>	<b>-1.622</b>	<b>-0.594</b>	<b>0.000004</b>	<b>0.583</b>	<b>0.042</b>	<b>1.124</b>	<b>0.028</b>
GN-RJ	<b>-1.074</b>	<b>-1.610</b>	<b>-0.538</b>	<b>0.000002</b>	0.154	-0.408	0.717	0.893
TT-EK	-0.173	-0.688	0.340	0.818	0.356	-0.183	0.897	0.322
GN-EK	-0.139	-0.675	0.396	0.906	-0.072	-0.635	0.490	0.987
GN-TT	0.033	-0.507	0.575	0.998	-0.429	-0.998	0.140	0.209
State								
During-Before	<b>0.454</b>	<b>0.040</b>	<b>0.867</b>	<b>0.027</b>	0.396	-0.037	0.830	0.081
After-Before	0.111	-0.301	0.524	0.799	0.102	-0.332	0.536	0.844
After-During	-0.342	-0.755	0.070	0.125	-0.294	-0.728	0.140	0.248

906

907  
908  
909  
910  
911  
912  
913  
914  
915  
916  
917  
918  
919  
920  
921  
922  
923  
924  
925  
926  
927  
928  
929  
930  
931  
932  
933  
934  
935  
936  
937  
938  
939  
940  
941  
942  
943  
944  
945  
946  
947

948 **Supplementary Table 8: Loadings of each behaviour in Principal Component (PC)**  
949 **composite variables for males in 14 minutes before and after calls in experiment 1**  
950

Behaviour	PC1	PC2	PC3
Rest	0.465	0.402	0.242
Fly	0.276	0.407	0.350
Bask	0.334	0.575	0.147
Flutter	0.443	0.378	0.235
Court	0.117	0.282	0.599
Copulate	0.006	0.093	0.039
Walk	0.489	0.275	0.321
Antenna wiggle	0.377	0.182	0.240
Sitting near each other	0.047	0.016	0.467
% Variance explained	29.23	19.82	15.88
% Total variance explained	29.23	49.06	64.94

951  
952  
953  
954  
955  
956  
957  
958  
959  
960  
961  
962  
963  
964  
965  
966  
967  
968  
969  
970  
971  
972

973 **Supplementary Table 9: Loadings of each behaviour in Principal Component (PC)**  
974 **composite variables for females in 14 minutes before and after calls in experiment 1**  
975  
976

Behaviour	PC1	PC2	PC3
Rest	0.548	0.285	0.204
Fly	0.320	0.272	0.104
Bask	0.469	0.416	0.153
Flutter	0.388	0.537	0.026
Copulate	0.045	0.136	0.842
Walk	0.376	0.446	0.051
Antenna wiggle	0.262	0.332	0.381
Sit near each other	0.041	0.087	0.246
Lifting abdomen	0.100	0.215	0.068
% Variance explained	28.99	17.63	12.24
% Total variance explained	28.99	46.63	58.87

977  
978  
979  
980  
981  
982  
983  
984  
985  
986  
987  
988  
989  
990  
991  
992  
993  
994  
995  
996  
997

998 **Supplementary Table 10: Effect of treatment (Rufous-tailed Jacamar, Eastern**  
 999 **Kingbird, Toco Toucan and Greenhouse background noise calls), state (before call**  
 1000 **and after call) and their interaction on 14 minute behaviours before and after call,**  
 1001 **and male PC1, PC2, PC3, courtship, copulation, sit near and female PC1, PC2, PC3,**  
 1002 **abdomen lifting behaviours in experiment 1.**  
 1003  
 1004

♂	AIC	Df	F value	Pr (>F)	♀	AIC	Df	F value	Pr (>F)
PC1	644				PC1	635			
Treatment		3	0.734	0.533	<b>Treatment</b>		<b>3</b>	<b>3.615</b>	<b>0.014</b>
State		1	0.014	0.907	State		1	1.488	0.224
Treatment*State		3	1.068	0.364	Treatment*State		3	0.236	0.871
PC2	580				PC2	560			
Treatment		3	1.482	0.222	Treatment		3	0.731	0.535
State		1	0.380	0.538	State		1	0.155	0.284
Treatment*State		3	0.133	0.952	Treatment*State		3	0.770	0.512
PC3	542				PC3	500			
Treatment		3	1.639	0.183	Treatment		3	1.142	0.334
State		1	0.664	0.416	State		1	0.498	0.481
Treatment*State		3	0.062	0.980	Treatment*State		3	0.433	0.730
Courtship	1606				Copulation	1983			
Treatment		3	1.313	0.272	Treatment		3	1.967	0.121
State		1	1.933	0.166	State		1	0	1
Treatment*State		3	0.243	0.866	Treatment*State		3	0	1
Sitting near other	2027				Abdomen lift	1676			
Treatment		3	0.953	0.417	Treatment		3	0.613	0.608
State		1	0.264	0.608	State		1	0.621	0.432
Treatment*State		3	0.221	0.882	Treatment*State		3	0.979	0.404



1005 **Supplementary Table 11: ANOVA post-hoc test results with PC as dependent**  
 1006 **variables and the treatments (calls), state (before and after) and their interaction as**  
 1007 **response variables for males and females in 14 minutes before and after calls in**  
 1008 **experiment 1. EK= Eastern kingbird; RJ=Rufous-tailed jacamar; TT=Toco toucan;**  
 1009 **GN=Greenhouse background noise; Difference= pairwise difference; lwr=lower**  
 1010 **range; upr=upper range; padj= adjusted p-value.**  
 1011

Treatment	Difference	♂ PC1			♀ PC1			
		lwr	upr	padj	Difference	lwr	upr	padj
EK-RJ	0.160	-0.755	1.076	0.968	0.064	-0.956	0.826	0.997
TT-RJ	0.305	-0.610	1.221	0.821	-0.043	-0.934	0.848	0.999
GN-RJ	0.528	-0.435	1.492	0.486	<b>0.948</b>	<b>0.010</b>	<b>1.887</b>	<b>0.046</b>
TT-EK	0.145	-0.759	1.050	0.975	0.021	-0.859	0.902	0.999
GN-EK	0.367	-0.586	1.321	0.748	<b>1.013</b>	<b>0.084</b>	<b>1.942</b>	<b>0.026</b>
GN-TT	0.222	-0.731	1.176	0.929	<b>0.992</b>	<b>0.063</b>	<b>1.920</b>	<b>0.031</b>
State								
After- Before	-0.029	-0.530	0.471	0.907	-0.301	-0.789	0.186	0.224
Treatment	Difference	PC2			PC2			
		lwr	upr	padj	Difference	lwr	upr	padj
EK-RJ	0.202	-0.552	0.956	0.898	0.177	-0.533	0.887	0.916
TT-RJ	-0.250	-1.005	0.503	0.823	<b>0.382</b>	<b>-0.328</b>	<b>1.093</b>	<b>0.502</b>
GN-RJ	-0.370	-1.165	0.423	0.620	0.305	-0.443	1.053	0.714
TT-EK	-0.452	-1.198	0.292	0.394	0.205	-0.497	0.907	0.872
GN-EK	-0.572	-1.358	0.213	0.235	0.128	-0.612	0.868	0.969
GN-TT	-0.119	-0.905	0.666	0.978	-0.077	-0.817	0.663	0.993
State								
After- Before	-0.128	-0.542	0.284	0.538	0.211	-0.177	0.600	0.284

1012  
 1013  
 1014

1015 **Supplementary Table 12: GLMM results on the effect of treatment (calls) and sex on**  
1016 **proportion of butterflies changing their behaviour at the start of calls in experiment**  
1017 **2. p<0.05 are bolded**

1018  
1019

Fixed effect	Estimate	SE	z value	Pr (> z )
<b>Intercept</b>	<b>-2.03</b>	<b>0.48</b>	<b>-4.18</b>	<b>&lt;0.0001</b>
Treatment (Eastern Kingbird)	0.57	0.57	1.01	0.312
<b>Treatment (Tropical Kingbird)</b>	<b>1.16</b>	<b>0.54</b>	<b>2.15</b>	<b>&lt;0.05</b>
Sex (male)	0.08	0.42	0.21	0.832
Random effect				
Order (Intercept)	6.9e-15	8.3e-8		

1020  
1021  
1022  
1023  
1024  
1025  
1026  
1027  
1028  
1029  
1030  
1031  
1032  
1033  
1034  
1035  
1036  
1037  
1038  
1039  
1040  
1041  
1042  
1043  
1044  
1045

1046 **Supplementary Table 13: Pairwise differences in the proportion of butterflies**  
1047 **changing their behavioural state in response to the start of calls in experiment 2**

1048  
1049

---

**ANOVA Type II Wald Chisquare test**

Treatment:  $\chi^2 = 4.807$ ;  $df = 2$ ;  $p\text{-value} = 0.09$

Sex:  $\chi^2 = 0.044$ ;  $df = 1$ ;  $p\text{-value} = 0.832$

---

**Pairwise comparisons between treatment**

Group 1	Group 2	p-value	Adj. p-value
Tropical Kingbird	Eastern Kingbird	0.336	1
Tropical Kingbird	Greenhouse noise	0.042	0.128
Eastern Kingbird	Greenhouse noise	0.402	1

1050  
1051  
1052  
1053  
1054  
1055  
1056  
1057  
1058  
1059  
1060  
1061  
1062  
1063

1064 **Supplementary Table 14: GLMM results of the effect of treatment (calls) and sex on**  
1065 **proportion of butterflies changing their behaviour at the end of calls in experiment**  
1066 **2. p<0.05 are bolded**

1067  
1068

Fixed effect	Estimate	SE	z value	Pr (> z )
<b>Intercept</b>	<b>-1.55</b>	<b>0.41</b>	<b>-3.70</b>	<b>&lt;0.001</b>
Treatment (Eastern Kingbird)	0.47	0.49	0.95	0.340
Treatment (Tropical Kingbird)	0.68	0.48	1.41	0.157
Sex (male)	0.07	0.39	0.19	0.844
Random effect				
Order (Intercept)	0	0		

1069  
1070  
1071  
1072  
1073  
1074  
1075  
1076  
1077  
1078  
1079  
1080  
1081  
1082  
1083  
1084  
1085  
1086  
1087  
1088  
1089  
1090  
1091  
1092  
1093  
1094

1095 **Supplementary table 15: Pairwise differences in the proportion of males and females**  
1096 **changing their behavioural state in response to the end of calls in experiment 2**

1097

1098

1099

---

**ANOVA Type II Wald Chisquare test**

Treatment:  $\chi^2 = 2.037$ ; df = 2; p-value = 0.361

Sex:  $\chi^2 = 0.038$ ; df = 1; p-value = 0.844

---

**Pairwise comparisons between treatment**

Group 1	Group 2	p-value	Adj. p-value
Tropical Kingbird	Eastern Kingbird	0.817	1
Tropical Kingbird	Greenhouse noise	0.231	0.693
Eastern Kingbird	Greenhouse noise	0.459	1

1100

1101

1102

1103

1104

1105

1106

1107

1108

1109

1110

1111

1112

1113

1114 **Supplementary Table 16: GLMM results of the effect of treatment (calls) and sex on**  
1115 **proportion of butterflies changing their behaviour in response to calls in experiment**  
1116 **2. p<0.05 are bolded**

1117  
1118

Fixed effect	Estimate	SE	z value	Pr (> z )
<b>Intercept</b>	<b>-1.31</b>	<b>0.38</b>	<b>-3.43</b>	<b>&lt;0.001</b>
<b>Treatment (Eastern Kingbird)</b>	<b>0.89</b>	<b>0.44</b>	<b>2.00</b>	<b>0.044</b>
<b>Treatment (Tropical Kingbird)</b>	<b>0.98</b>	<b>0.44</b>	<b>2.20</b>	<b>0.027</b>
Sex (male)	0.31	0.35	0.88	0.376
Random effect				
Order (Intercept)	0	0		

1119  
1120  
1121  
1122  
1123  
1124  
1125  
1126  
1127  
1128  
1129  
1130  
1131  
1132  
1133  
1134  
1135  
1136  
1137  
1138  
1139  
1140  
1141  
1142  
1143  
1144

1145 **Supplementary table 17: Pairwise differences in the proportion of individuals**  
1146 **changing their behavioural state in response to the calls in experiment 2**

1147  
1148

---

**ANOVA Type II Wald Chisquare test**

Treatment:  $\chi^2 = 5.756$ ;  $df = 2$ ;  $p\text{-value} = 0.056$

Sex:  $\chi^2 = 0.783$ ;  $df = 1$ ;  $p\text{-value} = 0.376$

---

**Pairwise comparisons between treatment**

Group 1	Group 2	p-value	Adj. p-value
Tropical Kingbird	Eastern Kingbird	1	1
Tropical Kingbird	Greenhouse noise	0.032	0.096
Eastern Kingbird	Greenhouse noise	0.052	0.158

1149  
1150  
1151  
1152  
1153  
1154  
1155  
1156  
1157  
1158  
1159  
1160  
1161  
1162  
1163

1164 **Supplementary Table 18: Loadings of each behaviour in Principal Component (PC)**  
1165 **composite variables for males in a minute before, during and after calls in**  
1166 **experiment 2**  
1167

Behaviour	PC1	PC2	PC3
Rest	0.537	0.514	0.139
Fly	0.292	0.017	0.413
Bask	0.274	0.401	0.400
Flutter	0.485	0.438	0.224
Court	0.258	0.026	0.332
Copulate	0.033	0.463	0.622
Walk	0.493	0.400	0.249
Antenna wiggle	0.056	0.074	0.194
% Variance explained	23.84	19.50	16.99
% Total variance explained	23.84	43.35	60.34

1168  
1169  
1170  
1171  
1172  
1173  
1174  
1175  
1176  
1177  
1178  
1179  
1180  
1181  
1182  
1183  
1184  
1185  
1186  
1187  
1188  
1189



1190 **Supplementary Table 19: Loadings of each behaviour in Principal Component (PC)**  
1191 **composite variables for females in a minute before, during and after calls in**  
1192 **experiment 2**  
1193  
1194

Behaviour	PC1	PC2	PC3
Rest	0.427	0.609	0.087
Fly	0.277	0.017	0.141
Bask	0.365	0.288	0.556
Flutter	0.506	0.314	0.319
Copulate	0.121	0.598	0.550
Walk	0.478	0.275	0.420
Antenna wiggle	0.307	0.045	0.278
Lifting abdomen	0.106	0.099	0.045
% Variance explained	27.03	19.39	15.16
% Total variance explained	27.03	46.42	61.58

1195  
1196  
1197  
1198  
1199  
1200  
1201  
1202  
1203  
1204  
1205  
1206  
1207  
1208  
1209  
1210  
1211  
1212  
1213  
1214  
1215

1216 **Supplementary Table 20: Effect of treatment (Tropical Kingbird, Eastern Kingbird,**  
 1217 **and Greenhouse background noise calls), state (before, during, and after call) and**  
 1218 **their interaction on male PC1, PC2, PC3, *courtship*, *sit near* and female PC1, PC2,**  
 1219 **PC3, *copulation*, *abdomen lifting* behaviours in experiment 2. p<0.05 bolded. For**  
 1220 **male *sit near* behaviour, there were zero occurrences.**  
 1221  
 1222

	♂	AIC	Df	F value	Pr (>F)	♀	AIC	Df	F value	Pr (>F)
PC1		760				PC1	784			
Treatment			2	0.062	0.940	Treatment		2	0.599	0.550
State			2	0.440	0.645	State		2	1.249	0.289
Treatment*State			4	0.249	0.910	Treatment*State		4	0.172	0.952
PC2		713				PC2	712			
Treatment			2	2.531	0.082	Treatment		2	2.361	0.096
State			2	0.667	0.514	State		2	0.207	0.813
Treatment*State			4	0.048	0.995	Treatment*State		4	0.167	0.954
PC3		683				PC3	661			
<b>Treatment</b>			<b>2</b>	<b>3.157</b>	<b>0.044</b>	Treatment		2	1.075	0.343
State			2	0.015	0.985	State		2	0.385	0.681
Treatment*State			4	0.205	0.935	Treatment*State		4	0.453	0.770
Courtship		1021				Copulation	1929			
Treatment			2	2.064	0.130	<b>Treatment</b>		<b>2</b>	<b>3.413</b>	<b>0.034</b>
State			2	0.292	0.747	State		2	0	1
Treatment*State			4	0.731	0.572	Treatment*State		4	0	1
Sitting near other		NA				Abdomen lift	1034			
Treatment			2	0	0	Treatment		2	1.279	0.280
State			2	0	0	State		2	1.588	0.207
Treatment*State			4	0	0	Treatment*State		4	1.292	0.274

1223 **Supplementary Table 21: ANOVA post-hoc test results with PC as dependent**  
 1224 **variables and the treatments (calls), state (before, during, and after) and their**  
 1225 **interaction as response variables for males and females in a minute before, during,**  
 1226 **and after calls in experiment 2. EK= Eastern kingbird; TK=Tropical kingbird;**  
 1227 **GN=Greenhouse background noise; Difference= pairwise difference; lwr=lower**  
 1228 **range; upr=upper range; padj= adjusted p-value.**  
 1229

		♂			♀			
		PC1			PC1			
Treatment	Difference	lwr	upr	padj	Difference	lwr	upr	padj
EK-TK	-0.055	-0.620	0.508	0.970	-0.120	-0.717	0.477	0.883
GN-TK	-0.080	-0.633	0.472	0.936	0.150	-0.435	0.735	0.817
GN-EK	-0.025	-0.578	0.527	0.993	0.270	-0.315	0.856	0.520
State								
During- Before	-0.213	-0.773	0.339	0.627	-0.078	-0.667	0.511	0.947
After- Before	-0.144	-0.701	0.411	0.812	-0.373	-0.962	0.215	0.294
After- During	0.072	-0.483	0.629	0.949	-0.295	-0.884	0.293	0.463
		PC2			PC2			
Treatment	Difference	lwr	upr	padj	Difference	lwr	upr	padj
EK-TK	-0.409	-0.914	0.095	0.136	0.028	-0.475	0.533	0.990
GN-TK	-0.417	-0.912	0.077	0.116	-0.376	-0.870	0.118	0.173
GN-EK	-0.007	-0.502	0.486	0.999	-0.404	-0.899	0.089	0.131
State								
During- Before	0.175	-0.322	0.673	0.684	-0.002	-0.499	0.495	0.999
After- Before	0.234	-0.263	0.731	0.508	0.116	-0.381	0.613	0.845
After- During	0.058	-0.438	0.556	0.957	0.118	-0.379	0.615	0.840

1230  
 1231

1232 **Supplementary Table 22: Loadings of each behaviour in Principal Component (PC)**  
1233 **composite variables for males in 14 minutes before and after calls in experiment 2**  
1234

Behaviour	PC1	PC2	PC3
Rest	0.013	0.435	0.374
Fly	0.325	0.102	0.225
Bask	0.047	0.233	0.631
Flutter	0.480	0.052	0.378
Court	0.538	0.207	0.190
Copulate	0.244	0.636	0.039
Walk	0.262	0.203	0.401
Antenna wiggle	0.014	0.455	0.217
Sit near each other	0.491	0.212	0.141
% Variance explained	24.60	20.21	17.82
% Total variance explained	24.60	44.81	62.64

1235  
1236  
1237  
1238  
1239  
1240  
1241  
1242  
1243  
1244  
1245  
1246  
1247  
1248  
1249  
1250  
1251  
1252  
1253  
1254  
1255  
1256

1257 **Supplementary Table 23: Loadings of each behaviour in Principal Component (PC)**  
1258 **composite variables for females in 14 minutes before and after calls in experiment 2**  
1259  
1260

Behaviour	PC1	PC2	PC3
Rest	0.210	0.708	0.151
Fly	0.350	0.029	0.221
Bask	0.430	0.234	0.262
Flutter	0.424	0.180	0.462
Copulate	0.278	0.603	0.319
Walk	0.452	0.090	0.429
Antenna wiggle	0.355	0.062	0.360
Sit near each other	0.078	0.175	0.468
Lifting abdomen	0.227	0.047	0.097
% Variance explained	29.96	18.61	13.99
% Total variance explained	29.96	48.58	62.58

1261  
1262  
1263  
1264  
1265  
1266  
1267  
1268  
1269  
1270  
1271  
1272  
1273  
1274  
1275  
1276  
1277  
1278  
1279  
1280  
1281

1282 **Supplementary Table 24: Effect of treatment (Rufous-tailed Jacamar, Eastern**  
 1283 **Kingbird, Toco Toucan and Greenhouse background noise calls), state (before call**  
 1284 **and after call) and their interaction on 14 minute behaviors before and after call,**  
 1285 **and male PC1, PC2, PC3, *courtship, copulation, sit near* and female PC1, PC2, PC3,**  
 1286 ***abdomen lifting* behaviours in experiment 2.**  
 1287  
 1288

♂	AIC	Df	F value	Pr (>F)	♀	AIC	Df	F value	Pr (>F)
PC1	527				PC1	555			
Treatment		2	0.558	0.574	Treatment		2	1.184	0.309
State		1	0.288	0.592	State		1	0.002	0.961
Treatment*State		2	0.840	0.434	Treatment*State		2	0.010	0.990
PC2	497				PC2	486			
Treatment		2	1.979	0.142	Treatment		2	1.363	0.259
State		1	1.212	0.273	State		1	0.463	0.497
Treatment*State		2	0.025	0.975	Treatment*State		2	0.280	0.756
PC3	483				PC3	443			
Treatment		2	0.009	0.991	Treatment		2	2.640	0.075
State		1	0.060	0.808	State		1	0.645	0.423
Treatment*State		2	0.391	0.677	Treatment*State		2	0.573	0.565
Courtship	1436				Copulation	2028			
Treatment		2	0.492	0.612	Treatment		2	2.821	0.063
State		1	0.332	0.565	State		1	0.838	0.361
Treatment*State		2	0.829	0.439	Treatment*State		2	0.112	0.894
Sitting near other	1148				Abdomen lift	1456			
Treatment		2	0.115	0.891	Treatment		2	0.101	0.904
State		1	1.041	0.309	State		1	0.002	0.966
Treatment*State		2	0.268	0.765	Treatment*State		2	0.554	0.576

1289 **Supplementary Table 25: ANOVA post-hoc test results with PC as dependent**  
 1290 **variables and the treatments (calls), state (before and after) and their interaction as**  
 1291 **response variables for males and females in 14 minutes before, during, and after**  
 1292 **calls in experiment 2. EK= Eastern kingbird; TK=Tropical kingbird;**  
 1293 **GN=Greenhouse background noise; Difference= pairwise difference; lwr=lower**  
 1294 **range; upr=upper range; padj= adjusted p-value.**  
 1295

Treatment	Difference	♂			♀			
		PC1			PC1			
		lwr	upr	padj	Difference	lwr	upr	padj
EK-TK	0.059	-0.683	0.802	0.980	-0.057	-0.879	0.763	0.984
GN-TK	0.304	-0.423	1.032	0.584	0.418	-0.387	1.223	0.437
GN-EK	0.244	-0.483	0.972	0.705	0.476	-0.329	1.281	0.343
State								
After- Before	-0.135	-0.634	0.363	0.592	0.013	-0.583	0.565	0.961
Treatment	Difference	♂			♀			
		PC2			PC2			
		lwr	upr	padj	Difference	lwr	upr	padj
EK-TK	-0.290	-0.958	0.378	0.560	-0.379	-1.024	0.264	0.345
GN-TK	-0.549	-1.204	0.105	0.118	-0.391	-1.023	0.239	0.308
GN-EK	-0.259	-0.914	0.395	0.615	-0.012	-0.643	0.619	0.998
State								
After- Before	-0.249	-0.698	0.199	0.272	-0.149	-0.582	0.284	0.497

1296